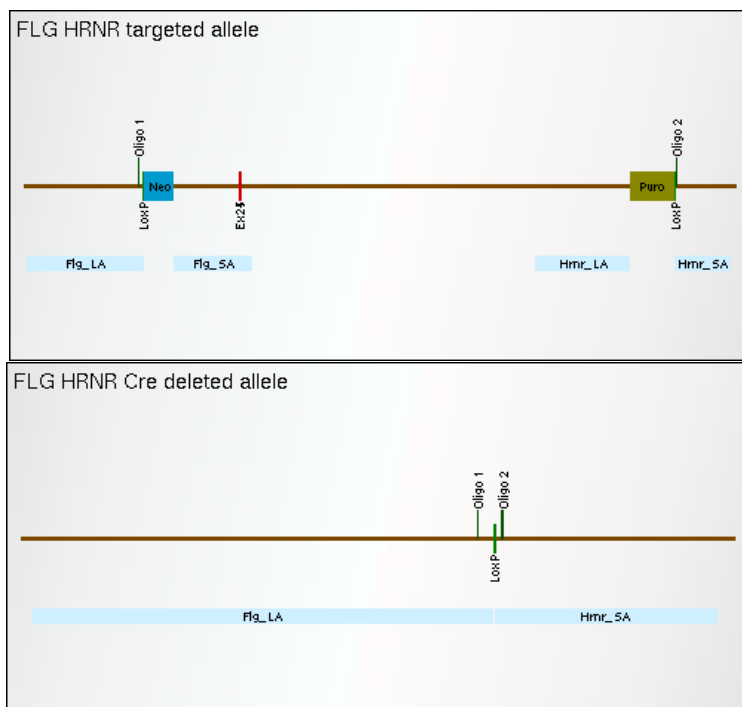


**EMMA ID: 05513**

**Gene: *Flg***

**Common name: *FLG KO 3 a conv***

### Allele Information



### Genotyping Information

Genotyping by end-point PCR based on gel is composed of a genespecific short range PCR using primers on wild type allele and a mutant allele-specific short range PCR. The combined results show the genotype of the mice. For example: mutant positive, wild type positive = Heterozygous.

#### PCR primer pairs and expected size bands

Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Control band	1260_1	1260_2	585
Mutant	1648_42	1648_44	426

#### Primer sequences

Primer Name	Sequence 5' --> 3'
1648_42	AGGCATGGTGGAACTGATGG
1648_44	TGTCCACAGTTAGATGACTG
1260_1	GAGACTCTGGCTACTCATCC
1260_2	CCTTCAGCAAGAGCTGGGGAC

### PCR setup (Qiagen, Hot Start Plus)

Component	Volume (µl) 1x	Final conc.
DNA (~ 50-100 ng)	2	
Q-Solution (5x)	2,5	0,5
PCR-Buffer (10x)	2,5	1
DNTP mix (10 mM)	0,5	0,2
MgCl <sub>2</sub> (25 mM)	1,5	1,5
Primer 1 (10 pmol/µl)	1	0,4
Primer 2 (10 pmol/µl)	1	0,4
Taq Polymerase (5 U/µl)	0,3	0,06
H <sub>2</sub> O*	13,7	
Final volume	25	

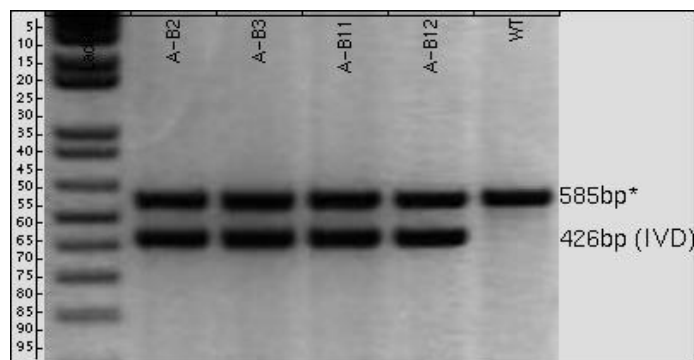
\* The amount of H<sub>2</sub>O is adjusted with the number of primer.

### Amplification conditions

PCR Settings	Temperature (°C)	Time	# of cycles
1 Denaturation (Melting)	95°C	5 min	1
2 Amplification (Melting, Annealing, Polym.)	94°C	30 sec	39
	60°C	45 sec	
	72°C	45 sec	
3 Polymerisation	72°C	10 min	1
4 Cooling	4°C	hold	1

These PCR conditions have been optimized for our methods and preparation kits. Adaptions may be required.

### Gel Image



**GSF IVD C-A1 specifically detects correct in vitro deletion in all subclones from C-A1 doubly targeted clone (\*=internal control for PCR amplification, shows presence of DNA template in all samples)**

Separated by gel electrophoresis on a 2% agarose gel.