



Genotyping protocol

N°8

IR00003110 / E205

(ICS internal reference)

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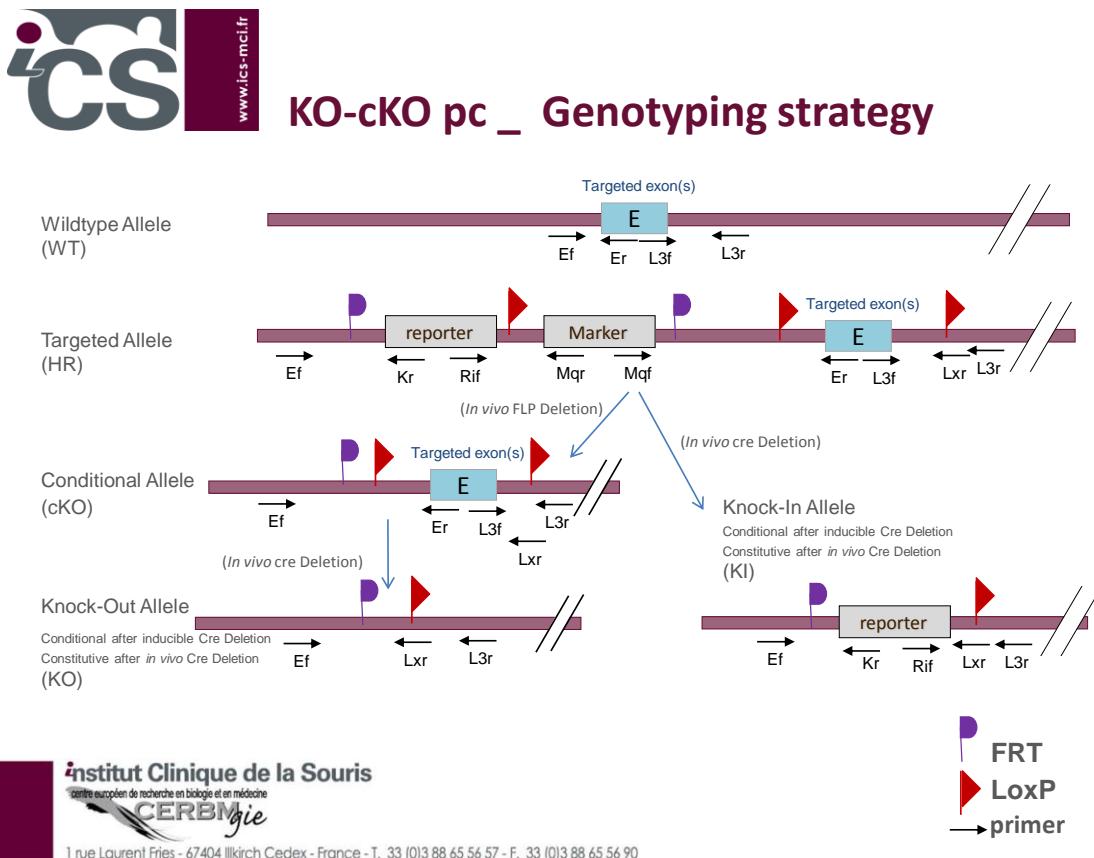
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1. Genotyping protocol and data

This section describes the condition used at the Mouse Clinical Institute (ICS) to genotype your **Nol8** Constitutive Knockout / Conditional Knockout (KO-cKO x Cre) project.

1.1. Genotyping strategy

The map below describes the position of the primers used for genotyping for each possible allele.



Sequence of primers used for genotyping:

Position	Primers	Sequence
Ef	5515	TGTGCCTGCTTGTAGTGATAATTGGA
Er	5513	TGTGGATATTAGGAACGTAACTGGGC
Kr	3277	CTCCTACATAGTTGGCAGTGTGGGG
L3f	5511	ACAAAACAAAATGGAAAGGTGGAACAC
L3r	5512	GAACCTGAAGCGACCTCTCACATC
Lxr	3254	TTATCATTAATTGCGTTCGCCATC
Lxr ²	3255	ACTGATGGCGAGCTCAGACCATAAC
Ri1f	5966	GCACATGGCTGAATATCGACGGT

²: for a selected position, a second primer was designed

PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer <i>(see the map above)</i>	Targeted allele (HR)	conditional allele (cKO)	KI allele	WildType allele
5' part of the selection marker	5515-3277	Ef / Kr	353	---	353	---
Presence of the distal loxP	5511-5512	L3f / L3r	476	476	---	449
Distal loxP specific PCR	5511-3254	L3f / Lxr	292	292	---	---
Excision of the selection marker	5515-5513	Ef / Er	7475*	519	---	333
Cre total excision	5966-3255	Ri1f / Lxr ²	---	---	471	---

*: this PCR product will not be observed using our PCR genotyping conditions (see description below)

**: this PCR is only verified if mice are generated

---: no Amplicon should be obtained

1.2. PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:

- FastStart PCR Master (Roche)
- DNA (50ng/ μ l)
- 5' primer (100 μ M)
- 3' primer (100 μ M)
- Sterile H₂O

Volume:

- 7.5 μ l
- 1.5 μ l
- 0.06 μ l
- 0.06 μ l
- up to 15 μ l

Cycling conditions:

Temp	Time	#Cycles
95°C	4min	1
94°C	30s	
62°C	30s	34
72°C	1min	
72°C	7min	1
20°C	5min	1

NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.

2. Cre and Flp genotyping method

You will find the genotyping protocol in the publication:

[Highly-efficient, fluorescent, locus directed cre and FlpO deleter mice on a pure C57BL/6N genetic background.](#)

Birling MC, Dierich A, Jacquot S, Hérault Y, Pavlovic G.
Genesis. 2012 Jun;50(6):482-9. doi:10.1002/dvg.20826. Epub 2012 Mar 20.