



## Genotyping protocol

Rnf144b

IR00003112 / E203

(ICS internal reference)

This report has been prepared by: **Christelle Morgenthaler**  
33 (0)3 88 65 56 55  
genotyping@igbmc.fr

This report has been validated by: **Sylvie Jacquot, PhD, Head of Genotyping Service**  
33 (0)3 88 65 57 44  
genotyping@igbmc.fr

The first version of this report was finalized the: 12 Dec 2013

For any question, please contact:

**Institut Clinique de la Souris - ICS - Mouse Clinical Institute**  
1 rue Laurent Fries, BP 10142  
67404 Illkirch Cedex, France  
Email: [genotyping@igbmc.fr](mailto:genotyping@igbmc.fr)  
Web site: <http://www-mci.u-strasbg.fr/>

## TABLE OF CONTENTS

**Table of contents** .....2

**1. Genotyping protocol and data** .....2

    1.1. Genotyping strategy .....2

    1.2. PCR protocol.....4

    1.3. Picture of genotyping with various alleles.....5

**2. Cre and Flp genotyping method**.....6

### 1. Genotyping protocol and data

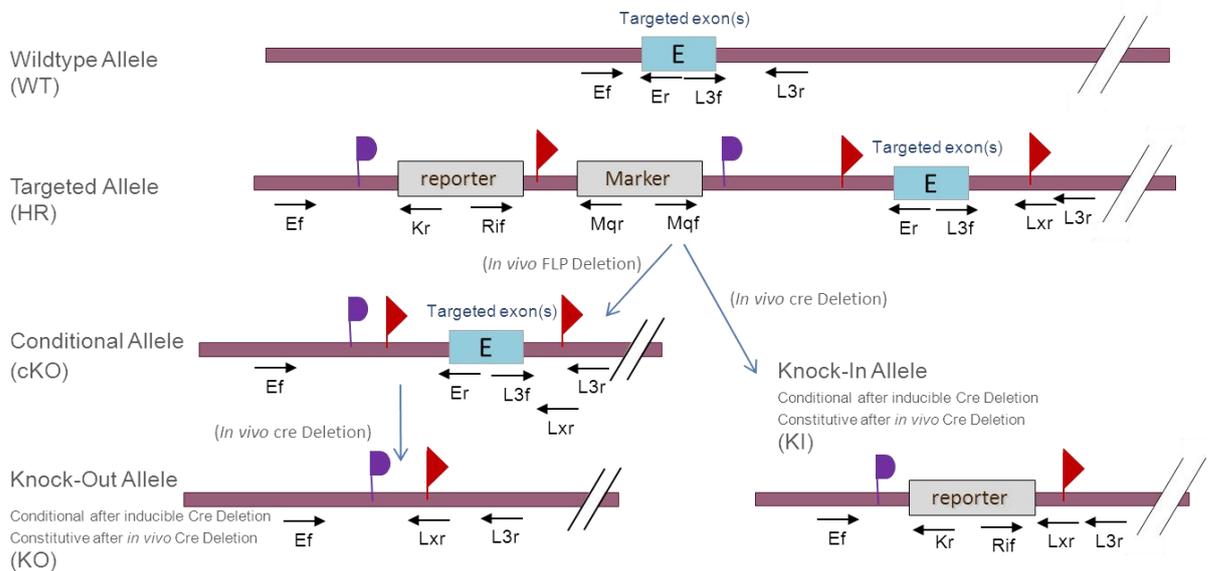
This section describes the condition used at the Mouse Clinical Institute (ICS) to genotype your **Rnf144b** Constitutive Knockout / Conditional Knockout (KO-cKO x Cre) project.

#### 1.1. Genotyping strategy

The map below describes the position of the primers used for genotyping for each possible allele.



### KO-cKO pc \_ Genotyping strategy



## Sequence of primers used for genotyping:

Position	Primers	Sequence
Ef	5507	GCTGTCTGAAGAGGACACAGGATGC
Ef <sup>2</sup>	5506	ACCGCATCACGTGTCATGTGGT
Er	5510	TCAGTGATGAAGTTCCTGATTGCC
Kr	3209	CCAACAGCTTCCCCACAACGG
L3f	5508	TCTCTACTGTCTTTGTTGCAGATTGC
L3r	5509	GCCTCGGCTGGTTAATGTTTGCA
Lxf	6295	TTATGTTTAAACGGCGCGCCC
Lxr	3255	ACTGATGGCGAGCTCAGACCATAAC
Ri1f	5966	GCACATGGCTGAATATCGACGGT

<sup>2</sup>: for a selected position, a second primer was designed

## PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer (see the map above)	Targeted allele (HR)	cKO allele	KI allele	WildType allele (WT)
5' part of the selection marker	5507-3209	Ef / Kr	339	---	---	---
Presence of the distal loxP	5508-5509	L3f / L3r	443	443	---	491
Distal loxP specific PCR	5508-3255	L3f / Lxr	313	313	---	---
Excision of the selection marker	5506-5510	Ef <sup>2</sup> / Er	7421*	465	---	298
Cre total excision	5966-3255	Ri1f / Lxr	3140*	---	474	---

\*: this PCR product will not be observed using our PCR genotyping conditions (see description below)

\*\* : this PCR is only verified if mice are generated

---: no Amplicon should be obtained

## 1.2. PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:	Volume:
- FastStart PCR Master (Roche)	7.5µl
- DNA (50ng/µl)	1.5µl
- 5' primer (100 µM)	0.06µl
- 3' primer (100 µM)	0.06µl
- Sterile H <sub>2</sub> O	up to 15 µl

### Cycling conditions:

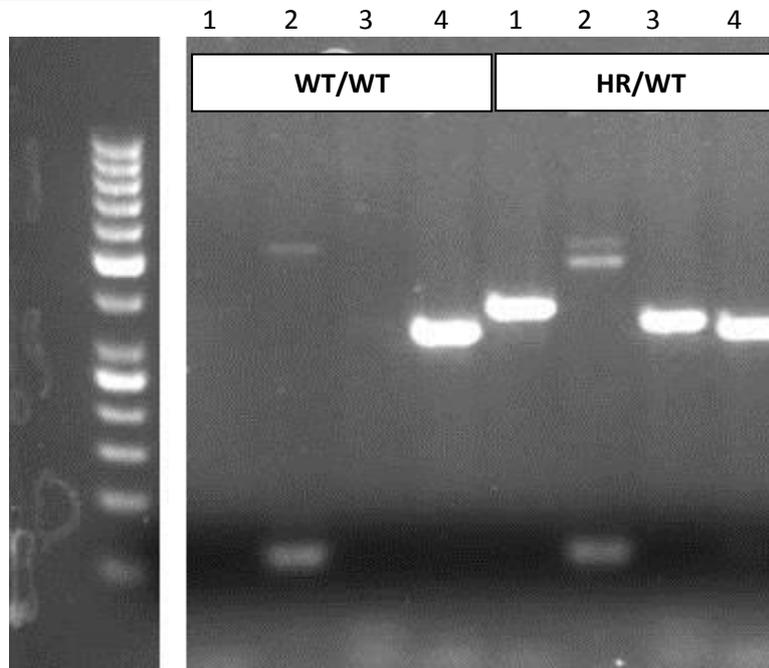
Temp	Time	#Cycles
95°C	4min	1
94°C	30s	34
62°C	30s	
72°C	1min	
72°C	7min	1
20°C	5min	1

**NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.**

### 1.3. Picture of genotyping with various alleles

Analysis of PCR products pattern was done by gel electrophoresis 2% agarose (SB buffer).

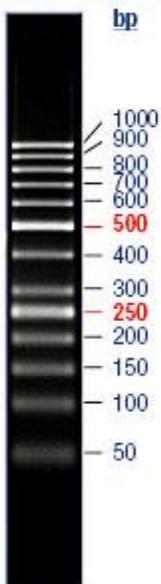
Representative genotyping picture



PCRs numbers:

- 1: 5' part of the selection marker
- 2: Presence of the distal loxP
- 3: Distal loxP specific PCR
- 4: Excision of the selection marker

#### O'GeneRuler™ 50bp DNA Ladder



## 2. Cre and Flp genotyping method

You will find the genotyping protocol in the publication:

[Highly-efficient, fluorescent, locus directed cre and FlpO deleter mice on a pure C57BL/6N genetic background.](#)

Birling MC, Dierich A, Jacquot S, Hérault Y, Pavlovic G.

Genesis. 2012 Jun;50(6):482-9. doi: 10.1002/dvg.20826. Epub 2012 Mar 20.