



Genotyping protocol

Clec9a KOMP

EPD0758_5_G08

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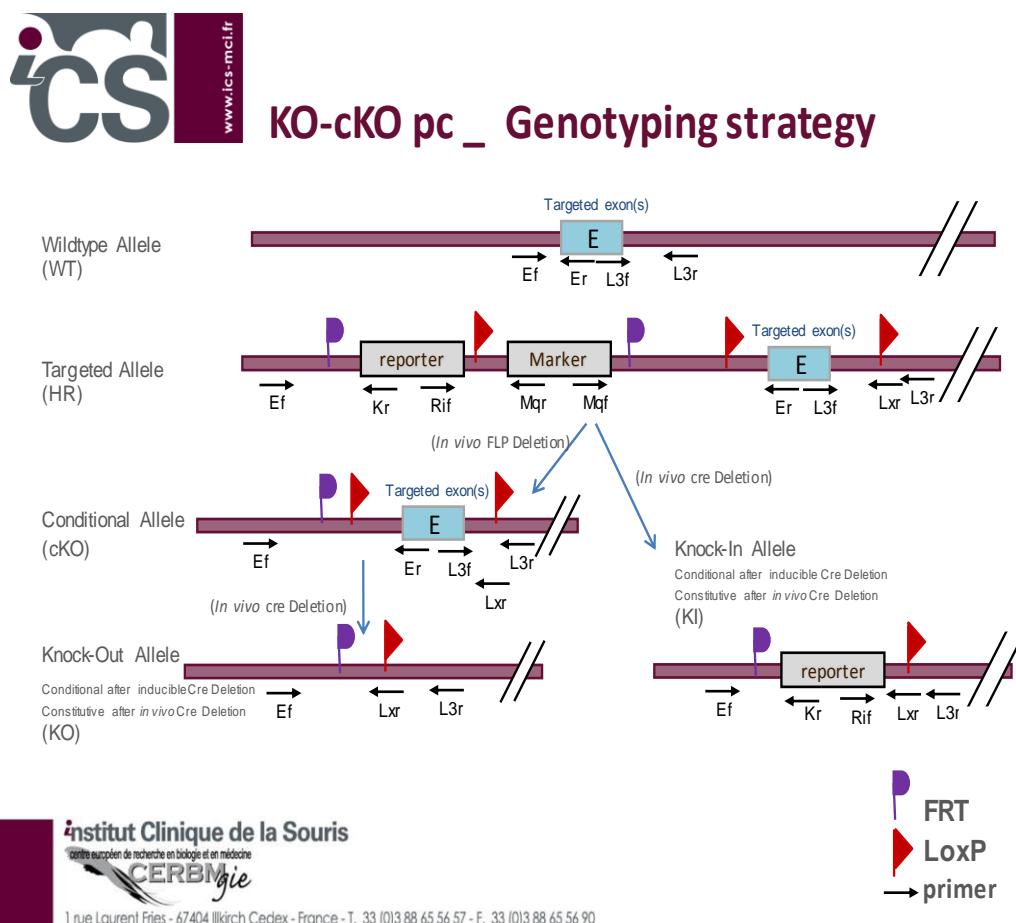
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1. Genotyping protocol and data

This section describes the condition used at the Mouse Clinical Institute (ICS) to genotype your **Clec9a** Constitutive Knockout / Conditional Knockout (KO-cKO x Cre) project.

1.1. Genotyping strategy

The map below describes the position of the primers used for genotyping for each possible allele.



Sequence of primers used for genotyping:

Position	Primers	Sequence
Ef	8106	CTCAGCCAATTCCATGCCTCAGTG
Ef ²	8107	CCATCCTTTGTGCATTAACGTGTTCTCTC
Er	8110	GCTTGAATTGGCTCAATTGCTGGTG
Kr	3210	CCTGTCCTCTCACCTTCTACC
L3f	8108	CTATGTCTTGAAACGCTGGGAAATGTG
L3r	8109	CATCCACAGTTGGGATGTACACGG
Lxr	3255	ACTGATGGCGAGCTCAGACCATAAC
Rif	5966	GCACATGGCTGAATATCGACGGT

²: for a selected position, a second primer was designed

PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer <i>(see the map above)</i>	Targeted allele (HR)	conditional allele (cKO)	KI allele	WildType allele
5' part of the selection marker (with Betaine)	8106-3210 (with 0.5% Betaine)	Ef / Kr	382	---	---	---
Presence of the distal loxP	8108-8109	L3f / L3r	436	436	---	402
Distal loxP specific PCR	8108-3255	L3f / Lxr	350	350	---	---
Excision of the selection marker (with Betaine)	8107-8110 (with 0.5% Betaine)	Ef ² / Er	7374*	470	---	285
Cre total excision	5966-3255	Rif / Lxr	3479*	---	471**	---

*: this PCR product will not be observed using our PCR genotyping conditions (see description below)

**: this PCR is only verified if mice are generated

---: no Amplicon should be obtained

1.2. PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:

- FastStart PCR Master (Roche)
- DNA (50ng/ μ l)
- 5' primer (100 μ M)
- 3' primer (100 μ M)
- Sterile H₂O

Volume:

- 7.5 μ l
- 1.5 μ l
- 0.06 μ l
- 0.06 μ l
- up to 15 μ l

Cycling conditions:

Temp	Time	#Cycles
95°C	4min	1
94°C	30s	
62°C	30s	34
72°C	1min	
72°C	7min	1
20°C	5min	1

NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.

2. Cre and Flp genotyping method

You will find the genotyping protocol in the publication:

[Highly-efficient, fluorescent, locus directed cre and FlpO deleter mice on a pure C57BL/6N genetic background.](#)

Birling MC, Dierich A, Jacquot S, Héault Y, Pavlovic G.
Genesis. 2012 Jun;50(6):482-9. doi:10.1002/dvg.20826. Epub 2012 Mar 20.