

EMMA ID: 09633

Gene: *Ech1*

Common name: *EPD0370_2_B11*

Allele: *Ech1*^{tm1a(EUCOMM)Wtsi}

Allele Information

Further information about the allele can be found on IMPC website at (copy the link to web browser)
[http://www.mousephenotype.org/data/alleles/MGI:1858208/tm1a\(EUCOMM\)Wtsi](http://www.mousephenotype.org/data/alleles/MGI:1858208/tm1a(EUCOMM)Wtsi)

Links to the general information

About IKMC resource

<https://www.infrafrontier.eu/knowledgebase/protocols/ikmc-products>

IKMC allele types

<http://www.i-dcc.org/kb/entry/89/>

Allele conversion guide - genotyping tm1b, tm1c and tm1d mice (assays infos available when required)

<http://www.mousephenotype.org/about-ikmc/targeting-strategies>

IMPC mouse phenotype data, search by the gene name

<http://www.mousephenotype.org/>

Genotyping Information

Genotyping by end-point PCR based on gel is composed of a genespecific short range PCR using primers on wild type allele and a mutant allele-specific short range PCR. The combined results show the genotype of the mice.

For example: mutant positive, wild type positive = Heterozygous.

PCR primer pairs and expected size bands

Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Mutant	Ech1 5' arm neu	LAR3	458
Wildtype	Ech1 5' arm neu	Ech1 3' arm neu	673

Primer sequences

Primer Name	Sequence 5' --> 3'
Ech1 5' arm neu	gctggcttaagaatgccagt
Ech1 3' arm neu	gttgagaaggatgagagggt
LAR3	CAACGGGTTCTTCTGTTAGTCC

PCR setup (Qiagen, Hot Start Plus)

Component	Volume (μ l) 1x	Final conc.
DNA (~ 50-100 ng)	2	
Q-Solution (5x)	2,5	0,5
PCR-Buffer (10x)	2,5	1
DNTP mix (10 mM)	0,5	0,2
MgCl ₂ (25 mM)	1,5	1,5
Primer 1 (10 pmol/ μ l)	1	0,4
Primer 2 (10 pmol/ μ l)	1	0,4
Taq Polymerase (5 U/ μ l)	0,3	0,06
H ₂ O*	13,7	
Final volume	25	

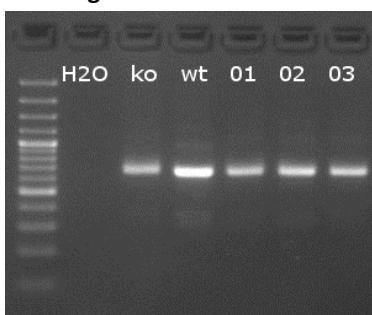
* The amount of H₂O is adjusted with the number of primer.

Amplification conditions

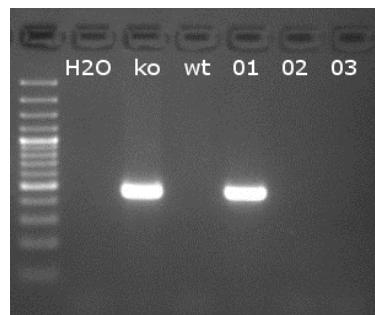
PCR Settings	Temperature (°C)	Time	# of cycles
1 Denaturation (Melting)	95°C	5 min	1
2 Amplification (Melting, Annealing, Polym.)	94°C 65°C 72°C	30 sec 45 sec 45 sec	39
3 Polymerisation	72°C	10 min	1
4 Cooling	12°C	hold	1

These PCR conditions have been optimized for our methods and preparation kits. Adoptions may be required.

Gel Image



WT-PCR



Mutant-PCR

Separated by gel electrophoresis on a 2% agarose gel.

WT-PCR doesn't work very well. High quality of DNA samples is suggested.

Genotyping using PCR-assays for cassette detection

LacZ reporter, Neo selection cassettes are inserted into the Knockout-first mutant allele. Cassette changes by allele conversion can be found on: <http://www.mousephenotype.org/about-ikmc/targeting-strategies>. For example, tm1b allele contains still lacZ reporter cassette, Neo selection cassette is deleted (promotor-driven only).

Please note that these assays are with universal cassette primers other than gene-specific. The confirmation on gene identity performed by e.g. sr genespecific PCR as provided is suggested .

PCR primer pairs and expected size bands

Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
lacZ	LacZ_multi_Deen_2F	LacZ_multi_Deen_2R	mut 81 bp,wt without band
Neo	Neo_long_Deen_F1	Neo_long_Deen_R1	mut 186 bp,wt without band

Primer sequences

Primer Name	Sequence 5' --> 3'
LacZ_multi_Deen_2F	TACTGGAGGCTGAAGTTCAGAT
LacZ_multi_Deen_2R	GCGTTTCACCCCTGCCATAA
Neo_long_Deen_F1	TTGAACAAGATGGATTGCACGC
Neo_long_Deen_R1	CCTCGTCCTGCAGTTCATT

PCR setup (Qiagen, Hot Start Plus)

Amplification conditions

Component	Volume (µl)	Final conc.	PCR Settings	Temperature (°C)	Time	# of cycles
DNA (~ 50-100 ng)	2		Denaturation (Melting)	95°C	5 min	1
Q-Solution (5x)	2,5	0,5	Amplification (Melting, An- nealing, Polym.)	94°C 58°C 72°C	30 sec 45 sec 45 sec	39
PCR-Buffer (10x)	2,5	1				
DNTP mix (10 mM)	0,5	0,2				
MgCl ₂ (25mM)	1,5	1,5	Polymerisation	72°C	10 min	1
Primer 1 (10 pmol/µl)	1	0,4				
Primer 2 (10 pmol/µl)	1	0,4				
Taq Polymerase (5 U/µl)	0,3	0,06				
H ₂ O	13,7		Cooling	12°C	hold	1
Final volume	25					

These PCR conditions have been optimized for our methods and preparation kits. Adoptions may be required.