



## Genotyping protocol

Ptpra

/ Ptpra

(ICS internal reference)

This report has been prepared by: **Christelle Roth**  
genotyping@igbmc.fr

This report has been validated by: **Sylvie Jacquot, PhD, Head of Genotyping Service**  
33 (0)3 88 65 57 44  
genotyping @igbmc.fr

The first version of this report was generated the: 17 Nov 2015

For any question, please contact:

**Institut Clinique de la Souris - ICS - Mouse Clinical Institute**

1 rue Laurent Fries, BP 10142

67404 Illkirch Cedex, France

Email: [genotyping@igbmc.fr](mailto:genotyping@igbmc.fr)

Web site: <http://www-mci.u-strasbg.fr/>

## TABLE OF CONTENTS

**Table of contents.....2**

**1. Genotyping protocol and data.....2**

    1.1. Genotyping strategy .....2

    1.2. PCR protocol.....4

    1.3. Picture of genotyping with various alleles .....5

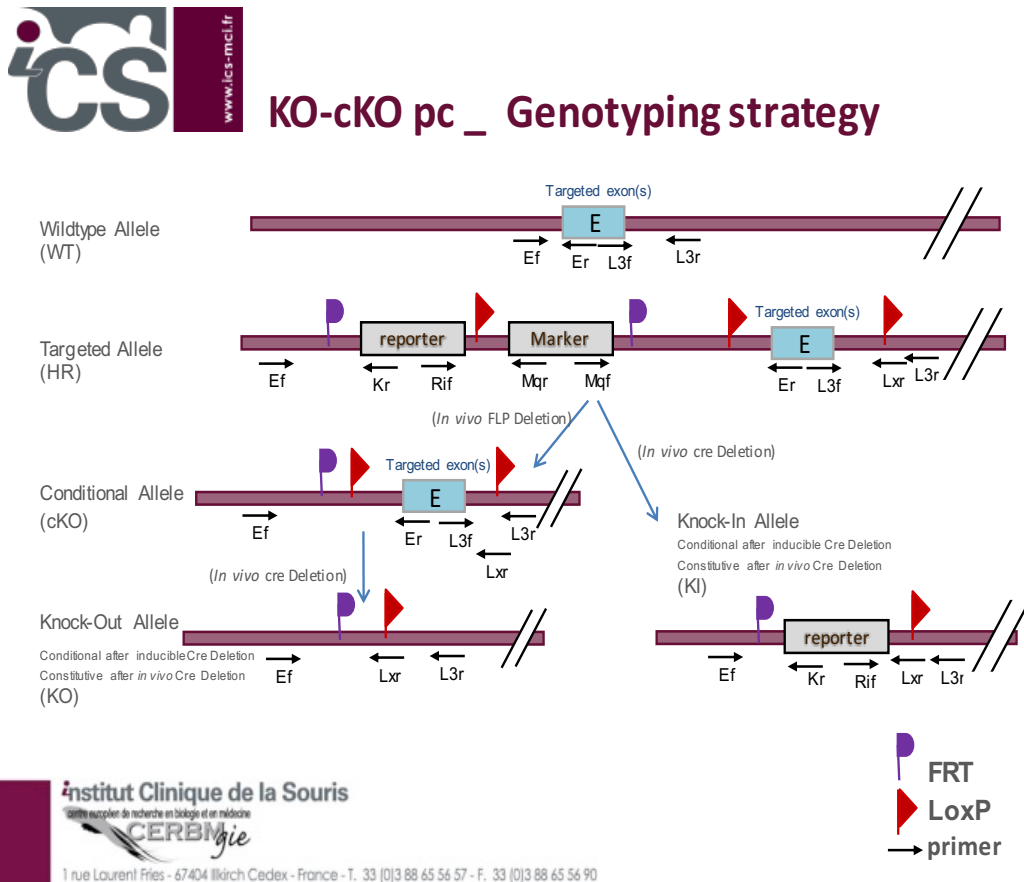
**2. Cre and Flp genotyping method .....6**

### 1. Genotyping protocol and data

This section describes the condition used at the Mouse Clinical Institute (ICS) to genotype your **Ptpra** Constitutive Knockout / Conditional Knockout (KO-cKO x Cre) project.

#### 1.1. Genotyping strategy

The map below describes the position of the primers used for genotyping for each possible allele.



### Sequence of primers used for genotyping:

Position	Primers	Sequence
Ef	Ef1	CTTTCCCTCCCTCTTCTGATTTTCTC
Kr	3277	CTCCTACATAGTTGGCAGTGTGGG
Kr	3209	CCAACAGCTCCCCACAACGG
L3f	L3f1	CTCTGATACTCCGGTGTCTCAGTAAGCC
L3f	L3f2	GGCCCTGTCCTCTCTGCTAGTAATCG
L3r	L3r1	CCTAAGACCAGCCTAGCAGACAAAGG
L3r	L3r2	GGACAAACCTGAAGATCTTGAAACACC
Lxr	3255	ACTGATGGCGAGCTCAGACCATAAC
Er	Er1	GCTAGAGTATGCGAGGAAAGAACCCTTG

<sup>2</sup>: for a selected position, a second primer was designed

### PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer (see the map above)	Targeted allele (HR)	conditional allele (cKO)	KI allele	WildType allele
5' part of the selection marker	Ef1-3277	Ef / Kr	271			
5' part of the selection marker	Ef2-3209	Ef / Kr	318			
Presence of the distal loxP	L3f1-L3r1	L3f/L3r	388	388		431
Presence of the distal loxP	L3f2-L3r2	L3f/L3r	349	349		391
Distal loxP specific PCR	L3f1-3255	L3f/Lxr	206	206		
Distal loxP specific PCR	L3f2-3255	L3f/Lxr	299	299		
Excision of the selection marker	Ef1/Er1	Ef/Er	7379	475		313
Cre total excision	5966-3255	Ri1f / Lxr	3175*	---	471**	

\*: this PCR product will not be observed using our PCR genotyping conditions (see description below)

\*\* : this PCR is only verified if mice are generated

---: no Amplicon should be obtained

## 1.2. PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:	Volume:
- FastStart PCR Master (Roche)	7.5µl
- DNA (50ng/µl)	1.5µl
- 5' primer (100 µM)	0.06µl
- 3' primer (100 µM)	0.06µl
- Sterile H <sub>2</sub> O	up to 15 µl

### Cycling conditions:

Temp	Time	#Cycles
95°C	4min	1
94°C	30s	34
62°C	30s	
72°C	1min	
72°C	7min	1
20°C	5min	1

**NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.**

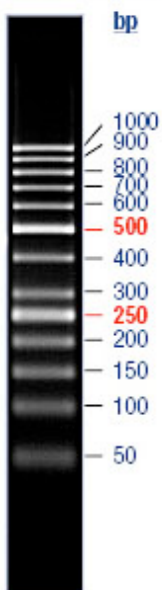
### 1.3. Picture of genotyping with various alleles

Analysis of PCR products pattern was done by gel electrophoresis 2% agarose (SB buffer).

Representative genotyping picture

\_ADD\_THE\_PHOTO\_

#### O'GeneRuler™ 50bp DNA Ladder



## 2. Cre and Flp genotyping method

You will find the genotyping protocol in the publication:

[Highly-efficient, fluorescent, locus directed cre and FlpO deleter mice on a pure C57BL/6N genetic background.](#)

Birling MC, Dierich A, Jacquot S, Hérault Y, Pavlovic G.

Genesis. 2012 Jun;50(6):482-9. doi: 10.1002/dvg.20826. Epub 2012 Mar 20.