

Genotyping protocol

Project Rr5865-DUP

(PHENOMIN-ICS reference None / Rr5865-DUP)

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1. Genotyping protocol and data

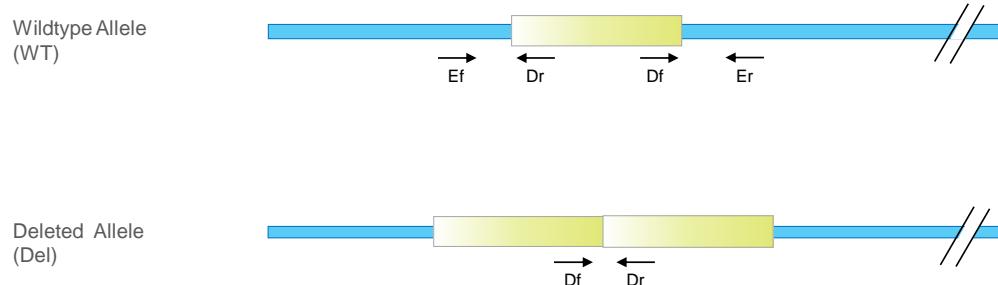
This section describes the condition used at the Mouse Clinical Institute (ICS) to genotype your Rr5865-DUP Crispr model, tandem duplication of genomic region (DUP) project.

1.1. Genotyping strategy

The map below describes the position of the primers used for genotyping for each possible allele.



Dup Genotyping strategy



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→ primer



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Sequence of primers used for genotyping:

Position	Primers	Sequence
Df	9100	AATGCTCTAACATATCCAGTGACC
Df ²	9183	TTACCAGAGTTATCCATGGGACACC
Df ³	9104	AAAGCATGACAGACGGGTAGTAAC
Dr	9105	ACACCTGAAAGGGGTTTACAGTG
Dr ²	9184	CCTGTCTCAGATTAGGATGCAGTC
Dr ³	9108	ACTGAACCCAGGGTCTGTATATGC
Ef	9099	CTCGTGGCTGATTGATTCTGTAAAG
Er	9106	CCAATGAACCATTGTCCTGCTATAC

^{2 (3)}: for a selected position, a second (third) primer was designed

PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer <i>(see the map above)</i>	Duplicated allele	WildType allele
WildType allele specific PCR (5' part of the targeted locus)	9099-9105	Ef / Dr	---	320
WildType allele specific PCR (3' part of the targeted locus)	9100-9106	Df / Er	---	390
PCR Dup	9104-9108	Df ³ / Dr ³	373	---
PCR_DUP2	9183-9184	Df ² / Dr ²	376	---

*: this PCR product will not be observed using our PCR genotyping conditions (see description below)

---: no Amplicon should be obtained



1.2. PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:	Volume:
- FastStart PCR Master (Roche)	7.5µl
- DNA (50ng/µl)	1.5µl
- 5' primer (100 µM)	0.06µl
- 3' primer (100 µM)	0.06µl
- Sterile H ₂ O	up to 15 µl

Cycling conditions:		
Temp	Time	#Cycles
95°C	4min	1
94°C	30s	
62°C	30s	34
72°C	1min	
72°C	7min	1
20°C	5min	1

NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.

2. Cre and Flp genotyping method

You will find the genotyping protocol in the publication:

[Highly-efficient, fluorescent, locus directed cre and FlpO deleter mice on a pure C57BL/6N genetic background.](#)

Birling MC, Dierich A, Jacquot S, Hérault Y, Pavlovic G.
Genesis. 2012 Jun;50(6):482-9. doi: 10.1002/dvg.20826. Epub 2012 Mar 20.



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