

**EMMA ID:** *EM:08175*  
**Gene:** *Pax6*  
**Common name:** *AEY080*  
**Allele:** *Pax6<sup>Aey80</sup>*

## Allele Information

Small eye mutant; ENU-induced;  
mutation affects Pax6 gene, intron 7: G->A at pos 1171

## Genotyping Information

Genotyping by end-point PCR based on gel is composed of a genespecific short range PCR using primers on wild type allele and a mutant allele-specific short range PCR. The combined results show the genotype of the mice. For example: mutant positive, wild type positive = Heterozygous. In addition to the expected product, the mutant assay may also amplify the endogenous wild type sequence, which will appear as a larger band on an agarose gel. The presence of this extra band will depend on the size of the original deletion.

### PCR primer pairs and expected size bands

Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Mutant	Pax6-Ex7-Int7-L1	Pax6-Ex7-Int7-R1	320

### Primer sequences

Primer Name	Sequence 5' --> 3'
Pax6-Ex7-Int7-L1	ACAGAGTTCTTCGCAACCTGGC
Pax6-Ex7-Int7-R1	CCTTGACATACATAATCCTTACAGTCACC

Sequencing of the PCR product

**PCR setup (AllTaq)**

Component	Volume (µl) 1x	Final conc.	
DNA (~ 50-100 ng)		2	
Q-Solution (5x)		2,5	0,5
PCR-Buffer (5x)		5	1
DNTP mix (10 mM)		0,5	0,2
MgCl <sub>2</sub> (25 mM)		1,5	1,5
Primer 1 (10 pmol/µl)		1	0,4
Primer 2 (10 pmol/µl)		1	0,4
Primer 3 (10 pmol/µl)		1	0,4
Taq Polymerase (5 U/µl)		0,5	2,5 U/rxn
H <sub>2</sub> O*		10	
Final volume		25	

\* The amount of H<sub>2</sub>O is adjusted with the number of primer.

**Amplification conditions**

PCR Settings	Temperature (°C)	Time	# of cycles
1 Denaturation (Melting)	95°C	5 min	1
2 Amplification (Melting, Annealing, Polym.)	94°C	30 sec	39
	67°C	45 sec	
	72°C	45 sec	
3 Polymerisation	72°C	10 min	1
4 Cooling	4°C	hold	1