

EMMA ID: *EM:13831*

Gene: *Tnfrsf19*

Common name: *Troy-CreERT2*

Allele: *Tnfrsf19^{tm1.1(cre/ERT2)Cle}*

Allele Information

The Troy-eGFP-IRES-CreERT2 transgenic mice express the fluorescence marker eGFP and the tamoxifen inducible recombinase (CreERT2) driven by the *Tnfrsf19* (Troy) promoter. Mice were generated by homologous recombination by inserting an eGFP-IRES-CreERT2 cassette with a Neomycin cassette flanked by loxP sites into the translational start site (exon 2) of *Tnfrsf19*.

Genotyping Information

Genotyping by end-point PCR based on gel is composed of a genespecific short range PCR using primers on wild type allele and a mutant allele-specific short range PCR. The combined results show the genotype of the mice. For example: mutant positive, wild type positive = Heterozygous. In addition to the expected product, the mutant assay may also amplify the endogenous wild type sequence, which will appear as a larger band on an agarose gel. The presence of this extra band will depend on the size of the original deletion.

PCR primer pairs and expected size bands

Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Wild type	<i>Tnfrsf19_gt_kirev1</i>	<i>Tnfrsf19_gt_wtrev1</i>	267
Mutant	<i>Tnfrsf19_gt_kirev1</i>	<i>Tnfrsf19_gt_for1</i>	314

Primer sequences

Primer Name	Sequence 5' --> 3'
<i>Tnfrsf19_gt_kirev1</i>	GCTGAACCTTGTGGCCGTTTAC
<i>Tnfrsf19_gt_for1</i>	GGCAAAACCCCAACACTTA
<i>Tnfrsf19_gt_wtrev1</i>	AGAATGGCAGCGAAGAGCAC

Component	Volume (µl) 1x	Final conc.
DNA (~ 50-100 ng)	2	
Q-Solution (5x)	2,5	0,5
PCR-Buffer (5x)	5	1
DNTP mix (10 mM)	0,5	0,2
MgCl ₂ (25 mM)	1,5	1,5
Primer 1 (10 pmol/µl)	1	0,4
Primer 2 (10 pmol/µl)	1	0,4
Primer 3 (10 pmol/µl)	1	0,4
Taq Polymerase (5 U/µl)	0,5	2,5 U/rxn
H ₂ O*	10	
Final volume	25	

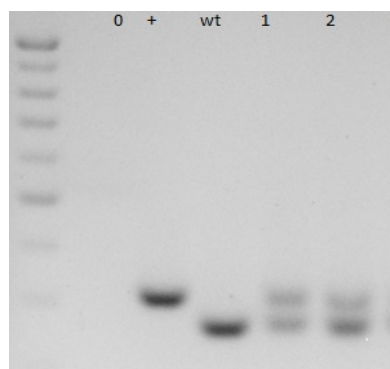
* The amount of H₂O is adjusted with the number of primer.

Amplification conditions

PCR Settings	Temperature (°C)	Time	# of cycles
1 Denaturation (Melting)	95°C	5 min	1
2 Amplification (Melting, Annealing, Polym.)	94°C	30 sec	39
	60°C	45 sec	
	72°C	45 sec	
3 Polymerisation	72°C	10 min	1
4 Cooling	4°C	hold	1

Touch-Down cycling protocol: first 10 cycles anneal at 65°C, decreasing 1°C per cycle, next 30 cycles anneal at 55°C. These PCR conditions have been optimized for our methods and preparation

Gel Image



+ hom
1,2 het

Separated by gel electrophoresis on a 2% agarose gel.