

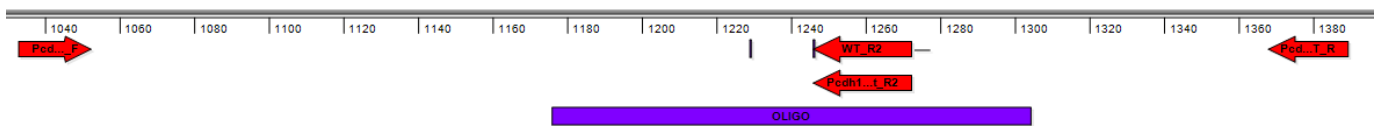
**Gene:** Pcdh15

**Colony prefix:** DAEX

**Allele:** Pcdh15<sup>em2Wtsi</sup>

**Allele type:** Point mutation

**Allele information:**



#### Mouse QC information

|                              |      |                              |      |
|------------------------------|------|------------------------------|------|
| Loss of WT Allele (LOA) qPCR | na   | Mutation Sequence confirmed  | pass |
| Mutant Specific SR-PCR       | pass | Off-target analysis complete | na   |

#### Guide RNAs used in initial experiment

| Sequence   | Chr | Chr Start | Chr End  |
|--|-----|-----------|----------|
| TCCTCTGAGAATTGTAGCTCTGG  | 10  | 74317130  | 74317152 |
| AGGATAAACACATAAGATAATCTCACCAAATAATTGCTAAGAT<br>AGTCATGGAACCTTACGGCTTCTATGTCTTTGTCTAGAGCTACAA<br>TTCTCAGAGGAGTGGTTAAGTTAGGCTCTCAGAAATGGTGGC | 10  | 74317098  | 74317227 |

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### Mutant allele sequence:

GCCACCATTTCTGAGAGCCTAAACTTAACCACTCCTCTGAGAATTGTAGCTCT[G/A]GACAAAGACATAGAAG[A/C]CG  
TAAGTTCCATGACTATCTTAGCAATTATTTGGTGAGATTATCTTATGTGTTTATCC

### Genotyping by end-point PCR

#### PCRs primer pairs and expected size bands

| Assay Type   | Assay      | Forward Primer | Reverse Primer | Expected Size Band (bp) |
|--------------|------------|----------------|----------------|-------------------------|
| Standard PCR | Wild type  | Pcdh15_WT_F    | Pcdh15_WT_R2   | 240                     |
| Standard PCR | Mutant     | Pcdh15_WT_F    | Pcdh15_Mut_R2  | 240                     |
| Standard PCR | Screening* | Pcdh15_WT_F    | Pcdh15_WT_R    | 357                     |

\*The screening PCR flanks the SNP region and can be used for sequence verification of the allele. The PCR will not distinguish wildtype from mutant mice, however as a product will be amplified in all cases.

#### Primer sequences

| Primer Name   | Primer Sequence (5' > 3')   |
|---------------|-----------------------------|
| Pcdh15_WT_R2  | ttgctaagatagtcacggaacttacGT |
| Pcdh15_Mut_R2 | ttgctaagatagtcacggaacttacGG |
| Pcdh15_WT_F   | TTTTTCAGGCTGAGCAGGAC        |
| Pcdh15_WT_R   | gcactaagactggctatgctga      |

#### Reaction setup

| Reagent                   | µl   |
|---------------------------|------|
| DNA (~50-100 ng)          | 1    |
| 10x Buffer                | 2    |
| MgCl <sub>2</sub> (50 mM) | 0.6  |
| Platinum Taq (Invitrogen) | 0.2  |
| dNTPs (100 mM)            | 0.2  |
| Primer 1 (10 µM)          | 0.4  |
| Primer 2 (10 µM)          | 0.4  |
| ddH <sub>2</sub> O        | 15.2 |
| Total                     | 20   |

#### Amplification conditions

| Step | Conditions            | Time     |
|------|-----------------------|----------|
| 1    | 94°C                  | 5 min    |
| 2    | 94°C                  | 30 sec   |
| 3    | 58°C                  | 30 sec   |
| 4    | 72°C                  | 1:30 sec |
| 5    | Go to '2' + 34 cycles | -        |
| 6    | 72°C                  | 5 min    |
| 7    | 12°C                  | forever  |

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## Links to information and frequently asked questions

MGP mouse phenotype data:

<http://www.mousephenotype.org>

How the "critical" exon is decided:

<http://www.i-dcc.org/kb/entry/102/>

## Relevant publications

White, J.K., Gerdin, A.-K., Karp, N.A., Ryder, E., Buljan, M., Bussell, J.N., Salisbury, J., Clare, S., Ingham, N.J., Podrini, C., et al. (2013). Genome-wide Generation and Systematic Phenotyping of Knockout Mice Reveals New Roles for Many Genes. *Cell* 154, 452–464.

Mali P, Yang L, Esvelt KM, et al (2013) RNA-guided human genome engineering via Cas9. *Science* 339:823–6. doi: 10.1126/science.1232033

Jinek M, Chylinski K, Fonfara I, et al (2012) A programmable dual-RNA-guided DNA endonuclease in adaptive bacterial immunity. *Science* 337:816–21. doi: 10.1126/science.1225829

Cong L, Ran FA, Cox D, et al (2013) Multiplex genome engineering using CRISPR/Cas systems. *Science* 339:819–23. doi: 10.1126/science.1231143

Singh P, Schimenti JC, Bolcun-Filas E (2014) A Mouse Geneticist's Practical Guide to CRISPR Applications. *Genetics* genetics.114.169771–. doi: 10.1534/genetics.114.169771

Brandl C, Ortiz O, Röttig B, et al (2015) Creation of targeted genomic deletions using TALEN or CRISPR/Cas nuclease pairs in one-cell mouse embryos. *FEBS Open Bio* 5:26–35. doi: 10.1016/j.fob.2014.11.009

Zhou J, Wang J, Shen B, et al (2014) Dual sgRNAs facilitate CRISPR/Cas9 mediated mouse genome targeting. *FEBS J*. doi: 10.1111/febs.12735

Kraft K, Geuer S, Will AJ, et al (2015) Deletions, Inversions, Duplications: Engineering of Structural Variants using CRISPR/Cas in Mice. *Cell Rep*. doi: 10.1016/j.celrep.2015.01.016

Shen B, Zhang J, Wu H, et al (2013) Generation of gene-modified mice via Cas9/RNA-mediated gene targeting. *Cell Res* 23:720–3. doi: 10.1038/cr.2013.46

Wang H, Yang H, Shivalila CS, et al (2013) One-step generation of mice carrying mutations in multiple genes by CRISPR/Cas-mediated genome engineering. *Cell* 153:910–8. doi: 10.1016/j.cell.2013.04.025

Yang H, Wang H, Shivalila CS, et al (2013) One-Step Generation of Mice Carrying Reporter and Conditional Alleles by CRISPR/Cas-Mediated Genome Engineering. *Cell* 154:1370–1379. doi: 10.1016/j.cell.2013.08.022

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