

Name of Mouse model or mutation:

BACH2-G26K-EM1-B6

Description:

Point mutation model made using CRISPR/Cas9.

Type of mutation:

Point mutation: G26K

Sequence details

WT

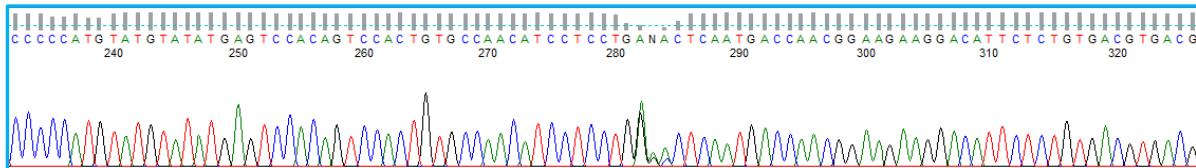
CAGCATATCCTTCAGCCGGTAACCAGGAAAACCTGTAGCTGCCCTGTCCCAGAGGAACAATAAT
ACAGCCACATTACTCAAAATCACACACATTAGCCTATTAGGCCTTCTAATTGTATCAGGAT
ATTAGCCCCAAAATTCCCGTGGATCCCACGTGTACAGAGTATGAGTCTCCTGCAGACTTTATTATACC
ATCTTTAAACCATTGATTAAAACCACACTTCTATTTCCTTCTTTCTTCTTCTTCTTCTTCTTCTTCTT
TCTTCTCTT
CACCTGAGTGCTGACTCATTCTGTTCGCAGCCCCACAGGAGAACATCAATATAAT
TAGCCCCAGGGCTATCCACAACACTACGGATGCTTAGTAGTCCTGCTCCCTGCCAGGGCTCATGG
GGACCTGGGAGCTTGGTGGCTTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTT
TTGAACGGCATGTCTGTGGATGAGAACGCTGGCTCCCCATGTATGTATATGAGTCCACAGTCCACT
GTGCCAACATCCTCCTGGCCTCAATGACCAACGGAAGAACGGACATTCTCTGTGACGTGACGCTGAT
CGTGGAGAGGAAGGAGTTCCGAGCCCACCGGGCTGTGCTGGCCGATGCAGTGAATACTCTGGCA
AGCACTGGTTGGACAGACGAAAGATGACTTGGTGGTCAGCTTGCCTGAGGAGGTACAGTGAATTGG
TGTGGTTGTGGGTTGTGGTGGAAAGTTAGACGTGCACTGTGCCACGTGGATGCCACGTACCCAT
GTCCACATCA

BACH2-G26K-EM1-B6

CAGCATATCCTTCAGCCGGTAACCAGGAAAACCTGTAGCTGCCCTGTCCCAGAGGAACAATAAT
ACAGCCACATTACTCAAAATCACACACATTAGCCTATTAGGCCTTCTAATTGTATCAGGAT
ATTAGCCCCAAAATTCCCGTGGATCCCACGTGTACAGAGTATGAGTCTCCTGCAGACTTTATTATACC
ATCTTTAAACCATTGATTAAAACCACACTTCTATTTCCTTCTTTCTTCTTCTTCTTCTTCTTCTTCTT
TCTTCTCTT
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TAGCCCCAGGGCTATCCACAACACTACGGATGCTTAGTAGTCCTGCTCCCTGCCAGGGCTCATGG
GGACCTGGGAGCTTGGTGGCTTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTTCTT
TTGAACGGCATGTCTGTGGATGAGAACGCTGGCTCCCCATGTATGTATATGAGTCCACAGTCCACT
GTGCCAACATCCTCCTG**AAA**CTCAATGACCAACGGAAGAACGGACATTCTCTGTGACGTGACGCTGAT
CGTGGAGAGGAAGGAGTTCCGAGCCCACCGGGCTGTGCTGGCCGATGCAGTGAATACTCTGGCA

AGCACTGGTTGGACAGACGAAAGATGACTTGGTGGTCAGCTGCCTGAGGAGGTACAGTGATTGG
TGTGGTTGTGGGTTGTGGTGGAAAGTTAGACGTGCACTGTGCCACGTGGATGCCACGTCACCCAT
GTCCACATCA

BACH2-G26K-EM1-B6 Heterozygous F1 animal sequence trace:



Nucleotide Alignment:

Predicted Protein Alignment:

| | | | | | | | |
|--------------|---------------------------|--|---|---|----|---|----|
| * | 20 | * | 40 | * | 60 | * | 80 |
| Bach2_WT : | MSVDEKPGSPMYVYESTVHCANILL | LNDQRKKDILCDVTLIVERKEFRAHRAVLAACSEYFWQALVGQTKDDLVLVSLPEEF | | | | | |
| Bach2_G26K : | MSVDEKPGSPMYVYESTVHCANILL | LNDQRKKDILCDVTLIVERKEFRAHRAVLAACSEYFWQALVGQTKDDLVLVSLPEEF | | | | | |
| | | MSVDEKPGSPMYVYESTVHCANILL | LNDQRKKDILCDVTLIVERKEFRAHRAVLAACSEYFWQALVGQTKDDLVLVSLPEEF | | | | |

QC strategy employed at Harwell to check the edited allele:

Genomic DNA was extracted from ear clip biopsies and amplified in a PCR reaction using the following conditions/primer sequences:

| | |
|-----------------------------------|--|
| Geno_BACH2_G26K_F3 primer (5'-3') | CAGCATATCCTCAGCCGGT |
| Geno_BACH2_G26K_R3 primer (5'-3') | TGATGTGGACATGGGTGACG |
| Taq Polymerase used | Thermofisher Super-Fi |
| Annealing Temperature (°C) | 61 |
| Elongation time (min) | 0.5 |
| WT product size (bp) | 891 |
| Mutant product size (bp) | 891 |
| Notes | Sequence with Geno_BACH2_G26K_F4 primer (5'-3': TAAGGTGGAAAGCAGGTTAC) and Geno_BACH2_G26K_R3 primer. |

All amplicons were sent for Sanger sequencing to check for integration of the donor oligo sequence at the target site. F1 sequences should be heterozygous unless on sex chromosome.

Copy counting by ddPCR

Copy counting of the donor sequence was carried out by ddPCR at the F1 stage to confirm donor oligos were inserted once on target into the genome. The following Taqman assay was used to copy count the donor sequence compared against a VIC-labelled reference assay for Dot1l:

| | |
|------------------------|-----------------------------|
| Assay name | BACH2-G26K-MUT1 |
| Forward Primer (5'-3') | ACTGTGCCAACATCCTCCTGAAA |
| Reverse Primer (5'-3') | GGTGGGCTCGGAACTCCTT |
| Probe (5'-3') | TGACCAACGGAAGAAGGACATTCTCTG |
| Label | FAM-BHQ1 |

The ddPCR assay is specific to the G26K mutation in the Bach2 gene and only the donor template sequence is expected to be recognised by this assay. WT controls are expected to call at 0 copies and a single integration for a correct mutation is expected to call at 1 copy for F1 (HET) animals.

| | |
|------------------------|-------------------------------|
| Assay name | BACH2-G26K-UNI1 |
| Forward Primer (5'-3') | CACTGTGCCAACATCCTCCT |
| Reverse Primer (5'-3') | GAAACTCCTCCTCTCCACGATCAG |
| Probe (5'-3') | CAACGGAAAGAAGGACATTCTCTGTGACG |
| Label | FAM-BHQ1 |

This ddPCR assay is universal to Bach2 WT and G26K alleles are recognised by this assay. WT controls are expected to call at 2 copies and a single integration for a correct mutation is expected to call at 2 copies for F1 (HET) animals.

| | |
|------------------------|---------------------------|
| Reference Assay Name | Dot1l |
| Forward primer (5'-3') | GCCCCAGCACGACCATT |
| Reverse primer (5'-3') | TAGTTGGCATCCTTATGCTTCATC |
| Probe (5'-3') | CCCAACAGGCCTGGATTCTCAATGC |
| Label | VIC |

VIC-labelled reference assay for Dot1l gene.

No random insertions of donor sequence were detected in the animals taken forward to establish the colony.



Allele Description

This is a CRISPR/Cas9 induced mutation creating a series of point mutations; G26K in *BACH2*. The stock was generated at MRC Harwell via microinjection of CRISPR/Cas9 reagents into 1-cell stage embryos.

qPCR Copy Counting Genotyping Strategy

The genotyping strategy presented here has been optimized for reagents and conditions used by the Genotyping Core at MRC Harwell. To genotype animals, we recommend researchers validate the assay independently. PCR cycling temperature and times may require additional optimization based on the specific genotyping reagents used.

Samples are genotyped using qPCR copy counting with both a wild type and a mutant assay against a known reference assay (*Dot1l* on chromosome 10; 2 copies present). Samples for this line are genotyped using the following primers and probe:

- Wild type (WT) assay with probe and reverse primer binding to the WT bases mutated in the mutant allele.
- Mutant assay with probe and reverse primer binding to the G601R, F606Y and R609H point mutations.

For autosomal genes that have been targeted, the following results would be expected:

| Genotype of the Modified allele | WT Assay | Mutant Assay |
|---------------------------------|----------|--------------|
| Wildtype | 2 | 0 |
| Heterozygous | 1 | 1 |
| Homozygous mutant | 0 | 2 |



Bach2-G26K

MRC | Harwell

Bach2-G26K-WT1 assay (FAM labelled)

TTGAACGGCATGTCTGTGGATGAGAAGCCTGGCTCCCCATGTATGTATATGAGTCCACAGTCCAC
TGTGCCAACATCCTCCTGggCTCAATGACCAACGGAAAGAAGGACATTCTGTGACGTGACGCTGA

Lower case letters denote bases changed in the mutant allele.

Probe sequence is in bold and shaded grey.

Primer sequences are in bold and underlined.

| Oligo Bach2-G26K | 5' label | Sequence 5' → 3' | 3' label | Oligo Type |
|----------------------|----------|--------------------------------------|----------|-------------------|
| Bach2-G26K-UNI_F | n/a | <u>CGGCATGTCTGTGGATGAGAAG</u> | n/a | Universal Forward |
| Bach2-G26K-UNI_PROBE | FAM | ATGTATGTATATGAGTCCACAGTCC | ZEN/IBFQ | Universal Probe |
| Bach2-G26K-WT_R | n/a | <u>CCGTTGGTCATTGAGGCC</u> | n/a | Wild type Reverse |

Bach2-G26K-MUT1 assay (FAM labelled)

TTGAACGGCATGTCTGTGGATGAGAAGCCTGGCTCCCCATGTATGTATATGAGTCCACAGTCCAC
TGTGCCAACATCCTCCTGaaCTCAATGACCAACGGAAAGAAGGACATTCTGTGACGTGACGCTGA

Lower case letters denote bases changed in the mutant allele.

Probe sequence is in bold and shaded grey.

Primer sequences are in bold and underlined.

| Oligo Bach2-G26K | 5' label | Sequence 5' → 3' | 3' label | Oligo Type |
|----------------------|----------|--------------------------------------|----------|-------------------|
| Bach2-G26K-UNI_F | n/a | <u>CGGCATGTCTGTGGATGAGAAG</u> | n/a | Universal Forward |
| Bach2-G26K-UNI_PROBE | FAM | ATGTATGTATATGAGTCCACAGTCC | BHQ | Universal Probe |
| Bach2-G26K-MUT_R | n/a | <u>CTTCCGTTGGTCATTGAGTTCA</u> | n/a | Mutant Reverse |



Dot1l internal control (VIC labelled)

CTGATGGGTGAGATCCTACAGAGTCCCATTGCCACCATGTGTGCTACGCCTGAAATAAGCCTT**GCC**
CCAGCACGACCATTCAGGG**CCAGCTCTCAAGTCG**ACTGTAAG**ATGAAGCATAAGGATGCCA**ACTAACA
GAAAACGACTAGAGGGGAAAAGAACAAAGGAAACAGAAGACGCAGCACTCCGGCTCCCTGGGTTGCCAGT
CACCTATGA

| Oligo Bach2-G26K | 5' label | Sequence 5' → 3' | 3' label | Oligo Type |
|------------------|----------|---------------------------------|----------|------------|
| Dot1l_Foreward | n/a | <u>GCCCCAGCACGACCATT</u> | n/a | WT Forward |
| Dot1l_Probe | VIC | CCAGCTCTCAAGTCG | BHQ | WT Probe |
| Dot1l_Reverse | n/a | <u>TAGTTGGCATCCTTATGCTTCATC</u> | n/a | WT Reverse |

Probe sequence is in bold and shaded grey

Primer sequences are in bold and underlined

DNA extraction method

DNA is extracted from ear clips using Applied Biosystems Taqman Sample-to-SNP Kit and qPCR run using 1:10 dilution from the crude preparation.

qPCR master mix 1X

| | |
|---|----------|
| Applied Biosystems GTX Taqman master mix | 5 µl |
| Dot1l_Foreward (20 µM) | 0.225 µl |
| Dot1l_Reverse (20 µM) | 0.225 µl |
| Dot1l_Probe (5 µM) | 0.2 µl |
| FAM Assay (probe 5 µM & primers 15 µM each) | 0.3 µl |
| ddH2O | 1.55 µl |
| DNA (1:10 dilution of ABI Sample-to-SNP prep) | 2.5 µl |

Each sample is ran in technical duplicate. Seven WT and/or mutant controls are also included in duplicate along with non-template controls.

qPCR cycling conditions

qPCR instrument: Applied Biosystems 7500/7900 or ThermoFisher QuantStudio 7

95°C for 20 sec

Then 40 cycles of;

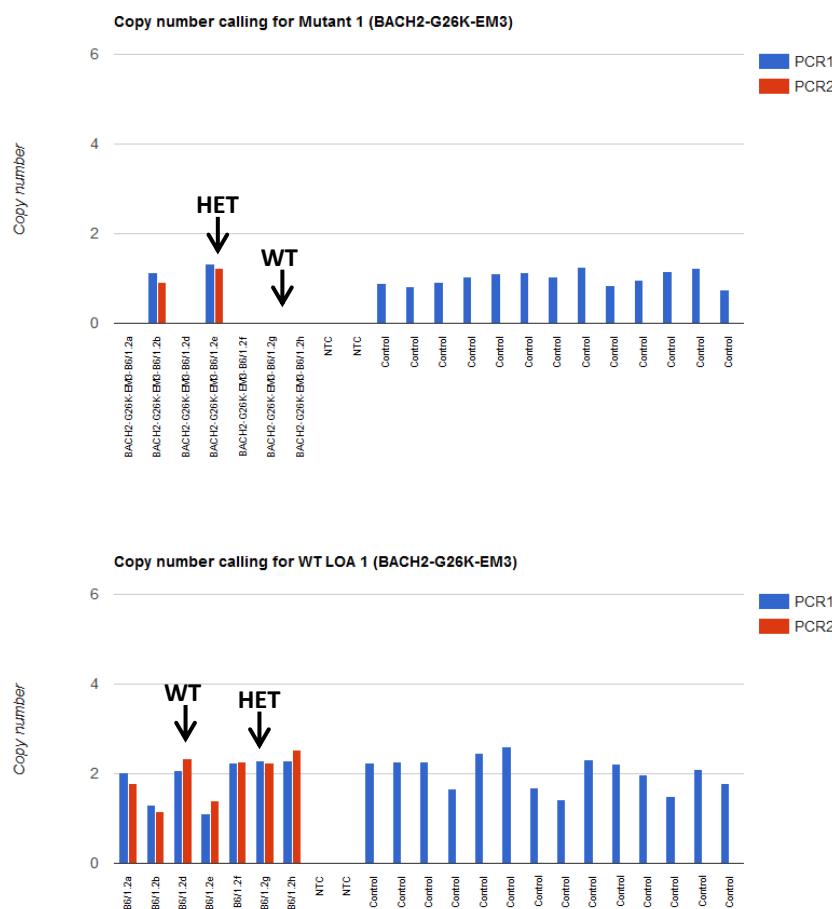
95°C for 3 sec

60°C for 30 sec

Analysis

The results are analysed using CopyCaller software v2.0 from Applied Biosystems or in-house software that is based on CopyCaller v2.0.

Bach2-G26K-WT1 and Bach2-G26K -MUT1 assays copy called results, image showing copy number chart for WT and Mutant assays (Task 280146 results)



Version No.

1

Date:

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Created/Updated by:

Daniel Ford

Approved by:

Rumana Zaman