

EMMA ID: 14847

Gene: Aox

Common name: Rosa26 Aox

Allele: *Gt(ROSA)26Sor^{tm1.1(CAG-AOX)Htj}*

Allele Information

Transgene: CAG-AOX-beta globin pA in Rosa 26 locus

Rosa26 construct integration



Dis Model Mech 2017 Szibor.pdf

Vector PL451 was adapted to serve as AOX entry vector, by integrating homology arms for the mouse Rosa26 locus (PL451-Rosa26) upstream and downstream of the neomycin selection cassette. Ciona AOX (Hakkaart et al., 2006) was integrated upstream of the selection cassette

Genotyping Information

Genotyping by end-point PCR based on gel is composed of a genespecific short range PCR using primers on wild type allele and a mutant allele-specific short range PCR. The combined results show the genotype of the mice. For example: mutant positive, wild type positive = Heterozygous. In addition to the expected product, the mutant assay may also amplify the endogenous wild type sequence, which will appear as a larger band on an agarose gel. The presence of this extra band will depend on the size of the original deletion.

PCR primer pairs and expected size bands

Assay	Forward Primer	Reverse Primer	Expected Size Band (bp)
Wild type	Rosa26 s	Rosa26 as	532
Mutant	Aox s	Aox as	317

Primer sequences

Primer Name	Sequence 5' --> 3'
Rosa26 s	GACCTCCATCGCGCACTCCG
Rosa26 as	CTCCGAGGCGGATCACAAGC
Aox s	GCGATGCAAGATGGAGGGTA
Aox as	TGAATCCAACCGTGGTCTCG

PCR setup (AllTaq)

Component	Volume (µl) 1x	Final conc.
DNA (~ 50-100 ng)	2	
Q-Solution (5x)	2,5	0,5
PCR-Buffer (5x)	5	1
DNTP mix (10 mM)	0,5	0,2
MgCl ₂ (25 mM)	1,5	1,5
Primer 1 (10 pmol/µl)	1	0,4
Primer 2 (10 pmol/µl)	1	0,4
Primer 3 (10 pmol/µl)	1	0,4
Taq Polymerase (5 U/µl)	0,5	2,5 U/rxn
H ₂ O*	10	
Final volume	25	

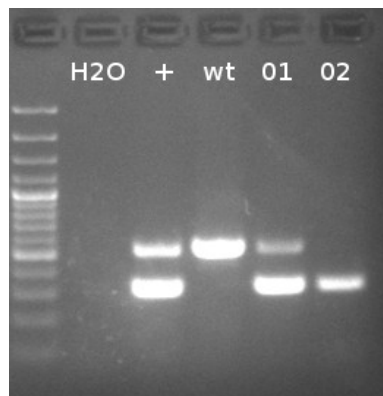
* The amount of H₂O is adjusted with the number of primer.

Amplification conditions

PCR Settings	Temperature (°C)	Time	# of cycles
1 Denaturation (Melting)	95°C	5 min	1
2 Amplification (Melting, Annealing, Polym.)	94°C 68-58 TD 72°C	30 sec 45 sec 45 sec	39
3 Polymerisation	72°C	10 min	1
4 Cooling	4°C	hold	1

Touch-Down cycling protocol: first 10 cycles anneal at 65°C, decreasing 1°C per cycle, next 30 cycles anneal at 55°C. These PCR conditions have been optimized for our methods and preparation

Gel Image



+ het control
1 het
2 hom

Separated by gel electrophoresis on a 2% agarose gel.