

OX40L-hCD4

Allele Description

This is a CRISPR/Cas9 induced knock-in of hCD4 Reporter into, ENSMUSE00000409027 of *OX40L*. The stock was generated at MRC Harwell via pronuclear injection of CRISPR/Cas9 reagents into 1-cell stage embryos.

qPCR Copy Counting Genotyping Strategy

The genotyping strategy presented here has been optimized for reagents and conditions used by the Genotyping Core at MRC Harwell. To genotype animals, we recommend researchers validate the assay independently. PCR cycling temperature and times may require additional optimization based on the specific genotyping reagents used.

Samples are genotyped using qPCR copy counting with both a wildtype loss of allele (WT-LOA) and a mutant assay against a known reference assay (*Dot1l* on chromosome 10; 2 copies present). Samples for this line are genotyped using the following primers and probe:

- Universal probe and Universal primer designed 5' of the deleted region.
- Wildtype specific primer situated within the deleted region.
- Mutant specific primer that binds to the inserted LoxP sequence

For autosomal genes that have been targeted, the following results would be expected:

| Genotype of the Modified allele | WT Assay | Mutant Assay |
|---------------------------------|----------|--------------|
| Wildtype | 2 | 0 |
| Heterozygous | 1 | 1 |
| Homozygous mutant | 0 | 2 |

OX40L-hCD4

OX40L-hCD4-WT1 assay (FAM labelled)

GGAAATTCAGATTAGTCACATAGAAGTTCCTCCCGCTGCAAAACCTGCAGAGTTAAAACGAAGAGA
CTCTGCTCTGCTCCTTCAATTGCTTTTTGTCTCCTGTTCTGGGACCTTTATCTTCTGACCCGCA**GGCTTG**
ACTTTGCCCTTATTGGCTCCTTTGTGGTGAAGAGCAGTCTTCCCCAGGTTCCCCGCCACAGCTGTAT
CTCCTCTGCACCC**CGACTGCAGAGATGGAAGG**GGAAGGGGTTCAACCCCTGGATGAGAATCTGGA
AAACGGATCAAGGCCAAGATTCAAGTGGAGAAGACGCTAAGGCTGGTGGTCTCTGGGATCAAGG

Lower case letters denote the deleted sequence in the mutant allele.

Probe sequence is in bold and shaded grey

Primer sequences are in bold and underlined

| Oligo OX40L-hCD4 | 5' label | Sequence 5' → 3' | 3' label | Oligo Type |
|----------------------|----------|-------------------------------------|----------|-------------------|
| OX40L-hCD4-UNI_F | n/a | <u>GGCTTGACTTTGCCCTTATTG</u> | n/a | Universal Forward |
| OX40L-hCD4-UNI_PROBE | FAM | AGACTGCTCTTACCACAAAGGAGC | ZEN-IBFQ | Universal Probe |
| OX40L-hCD4-WT_R | n/a | <u>CCTTCCATCTCTGCAGTCG</u> | n/a | WT Reverse |

OX40L-hCD4 -MUT1 assay (FAM labelled)

GGAAATTCAGATTAGTCACATAGAAGTTCCTCCCGCTGCAAAACCTGCAGAGTTAAAACGAAGAGA
CTCTGCTCTGCTCCTTCAATTGCTTTTTGTCTCCTGTTCTGGGACCTTTATCTTCTGACCCGCA**GGCTTG**
ACTTTGCCCTTATTGGCTCCTTTGTGGTGAAGAGCAGTCTTCCCCAGGTTCCCCGCCACAGCTGTAT
CTCCTCTGCACCCCGACTGCAGAGatgaaccgaggagtcctt**tttaggcacttgcttctggg**gctgcaactggcgctcctc
ccagcagccactcagggaaagaaagtgggctgggcaaaaaaggggatacagtgggaactgacctgtacagctcccagaagaag

Lower case letters denote the inserted sequence

Probe sequence is in bold and shaded grey

Primer sequences are in bold and underlined

| Oligo OX40L-hCD4 | 5' label | Sequence 5' → 3' | 3' label | Oligo Type |
|----------------------|----------|-------------------------------------|----------|-------------------|
| OX40L-hCD4-UNI_F | n/a | <u>GGCTTGACTTTGCCCTTATTG</u> | n/a | Universal Forward |
| OX40L-hCD4-UNI_PROBE | FAM | AGACTGCTCTTACCACAAAGGAGC | ZEN-IBFQ | Universal Probe |
| OX40L-hCD4-MUT_R | n/a | <u>ACCAGAAGCAAGTGCCTAAA</u> | n/a | Mutant Reverse |

OX40L-hCD4

Dot1l internal control (VIC labelled)

CTGATGGGTGTGGGCAGATCCTACAGAGTCCCATTGGCCACCATGTGTGCTACGCCTGAAATAAAGCCTT**GCC**
CCAGCACGACCATTCAGGG**CCAGCTCTCAAGTCG**ACTGTAAGATGAAGCATAAGGATGCCAACTACTAACA
GAAAACGACTAGAGGGGAAAAGAACAAGGAAACAGAAGACGCAGCACTCCGGCTTCCCTGGGTTGGCCAGT
CACCTATGA

| Oligo OX40L-hCD4 | 5' label | Sequence 5' → 3' | 3' label | Oligo Type |
|------------------|----------|--|----------|------------|
| Dot1l_Forward | n/a | <u>GCCCCAGCACGACCATT</u> | n/a | WT Forward |
| Dot1l_Probe | VIC | CCAGCTCTCAAGTCG | BHQ | WT Probe |
| Dot1l_Reverse | n/a | <u>TAGTTGGCATCCTTATGCTTCATC</u> | n/a | WT Reverse |

Probe sequence is in bold and shaded grey

Primer sequences are in bold and underlined

DNA extraction method

DNA is extracted from ear clips using Applied Biosystems Taqman Sample-to-SNP Kit and qPCR run using 1:10 dilution from the crude preparation.

qPCR master mix

1X

| | |
|---|----------|
| Applied Biosystems GTX Taqman master mix | 5 µl |
| Dot1l_Forward (20 µM) | 0.225 µl |
| Dot1l_Reverse (20 µM) | 0.225 µl |
| Dot1l_Probe (5 µM) | 0.2 µl |
| FAM Assay (probe 5 µM & primers 15 µM each) | 0.3 µl |
| ddH ₂ O | 1.55 µl |
| DNA (1:10 dilution of ABI Sample-to-SNP prep) | 2.5 µl |

Each sample is ran in technical duplicate. Seven WT and/or mutant controls are also included in duplicate along with non-template controls.

qPCR cycling conditions

qPCR instrument: Applied Biosystems 7500/7900 or ThermoFisher QuantStudio 7

95°C for 20 sec

Then 40 cycles of;

95°C for 3 sec

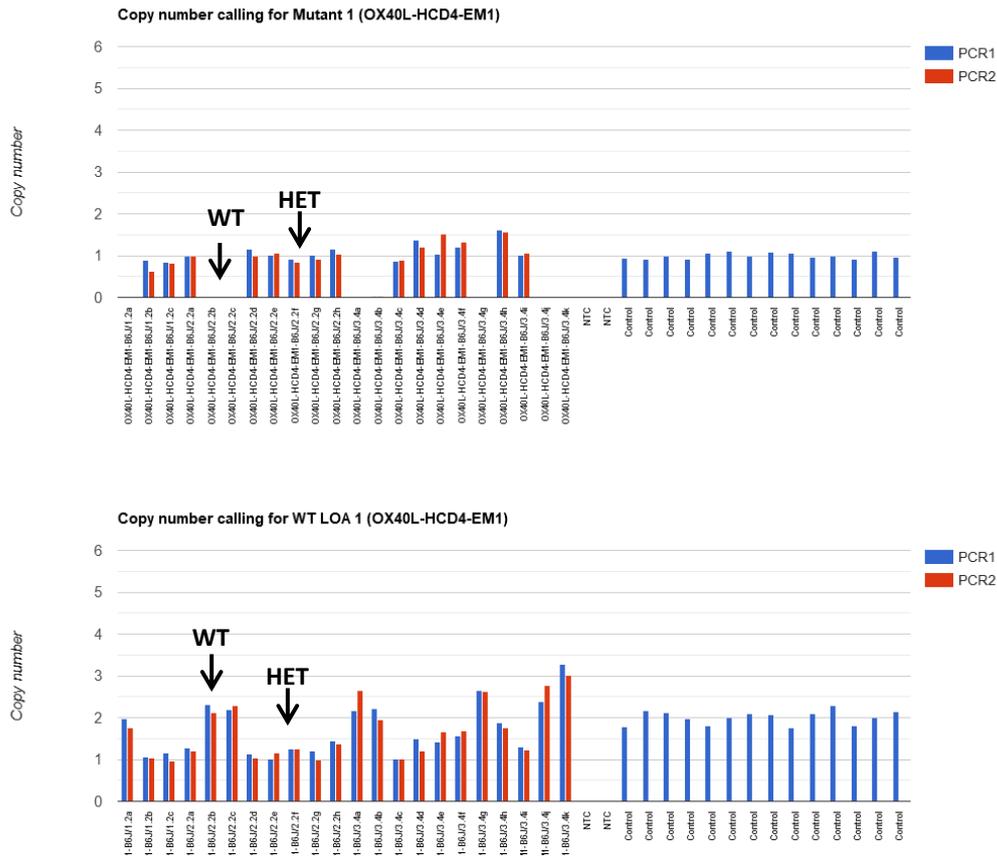
60°C for 30 sec

OX40L-hCD4

Analysis

The results are analysed using CopyCaller software v2.0 from Applied Biosystems or in-house software that is based on CopyCaller v2.0.

OX40L-hCD4'-WT1 and OX40L-hCD4-MUT1 assays copy called results, image showing copy number chart for WT and Mutant assays (Task 376432 results)



Version No. 1

Date: 03/08/2022

Created/Updated by: Daniel Ford

Approved by: Debbie Williams