

Genotyping protocol

Project mEos4 KI in Glrb

(PHENOMIN-ICS reference IR00006068 / Kos6068)

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1. Genotyping protocol and data

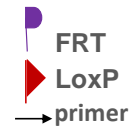
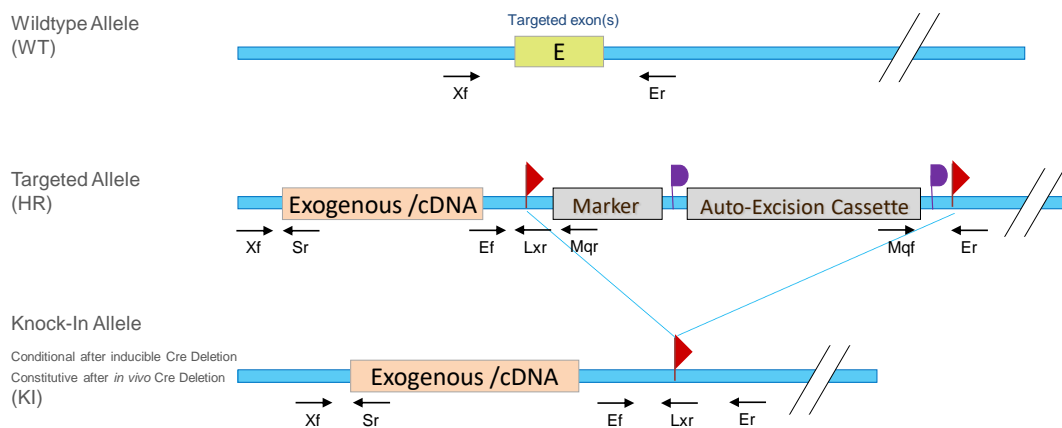
This section describes the condition used at the Mouse Clinical Institute (ICS) to genotype your **mEos4 KI in Glrb** Knockin (KI) project.

1.1. Genotyping strategy

The map below describes the position of the primers used for genotyping for each possible allele.



KI Genotyping strategy



Sequence of primers used for genotyping:

Position	Primers	Sequence
Ef	9202	TGCCCACCTACAGCACTGTTTGCAT
Er	9203	GCCCACAGCATCCAATTAAGTTGC
Lxr	4774	GAAGTTATACTAGAGCGCCGTTCCAC
Mq1f	1219	CAGCTCATTCTCCCACTCATGATC
Mq1r	265	TGCTAAAGCGCATGCTCCAGACTGC
Sr	9205	CGGAGTTTGATCCTCATGTCTGGCTT
Xf	9204	AGGACACTCAAACGTTTCAGTGATGTAGGAT

PCR fragments expected size (bp):

Region analyzed	Primers used	Position on the primer (see the map above)	Targeted allele (HR)	KI allele	WildType allele
Excision of the selection marker	9202-9203	Ef / Er	4445*	290	211
5' part of the selection marker	9202-265	Ef / Mq1r	297	---	---
3' part of the selection marker	1219-9203	Mq1f / Er	355	---	---
Exogenous/cDNA specific PCR	9204-9205	Xf / Sr	282	282	---
LoxP specific PCR	9202-4774	Ef / Lxr	205	205	---

*: this PCR product will not be observed using our PCR genotyping conditions (see description below)

---: no Amplicon should be obtained



1.2. PCR protocol

This section describes the composition of the mix and cycling conditions used for genotyping.

Reagents:	Volume:
- FastStart PCR Master (Roche)	7.5µl
- DNA (50ng/µl)	1.5µl
- 5' primer (100 µM)	0.06µl
- 3' primer (100 µM)	0.06µl
- Sterile H ₂ O	up to 15 µl

Cycling conditions:

Temp	Time	#Cycles
95°C	4min	1
94°C	30s	34
62°C	30s	
72°C	1min	
72°C	7min	1
20°C	5min	1

NB: These PCR conditions have been optimized for high-throughput genotyping. Adaptation to small-scale may be required.

2. Cre and Flp genotyping method

You will find the genotyping protocol in the publication:

[Highly-efficient, fluorescent, locus directed cre and FlpO deleter mice on a pure C57BL/6N genetic background.](#)

Birling MC, Dierich A, Jacquot S, Hérault Y, Pavlovic G.
Genesis. 2012 Jun;50(6):482-9. doi: 10.1002/dvg.20826. Epub 2012 Mar 20.

