



Introduction of a causative point mutation in Nr2f1 by CRISPR/Cas9 Genotyping protocol

Kus7772 / IM7772

Done by Dr Marie-Christine Birling, birlingm@igbmc.fr

24/08/2021

Strategy validated

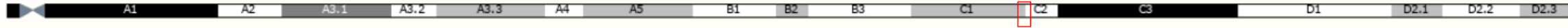


Nr2f1 Mouse Genomic locus

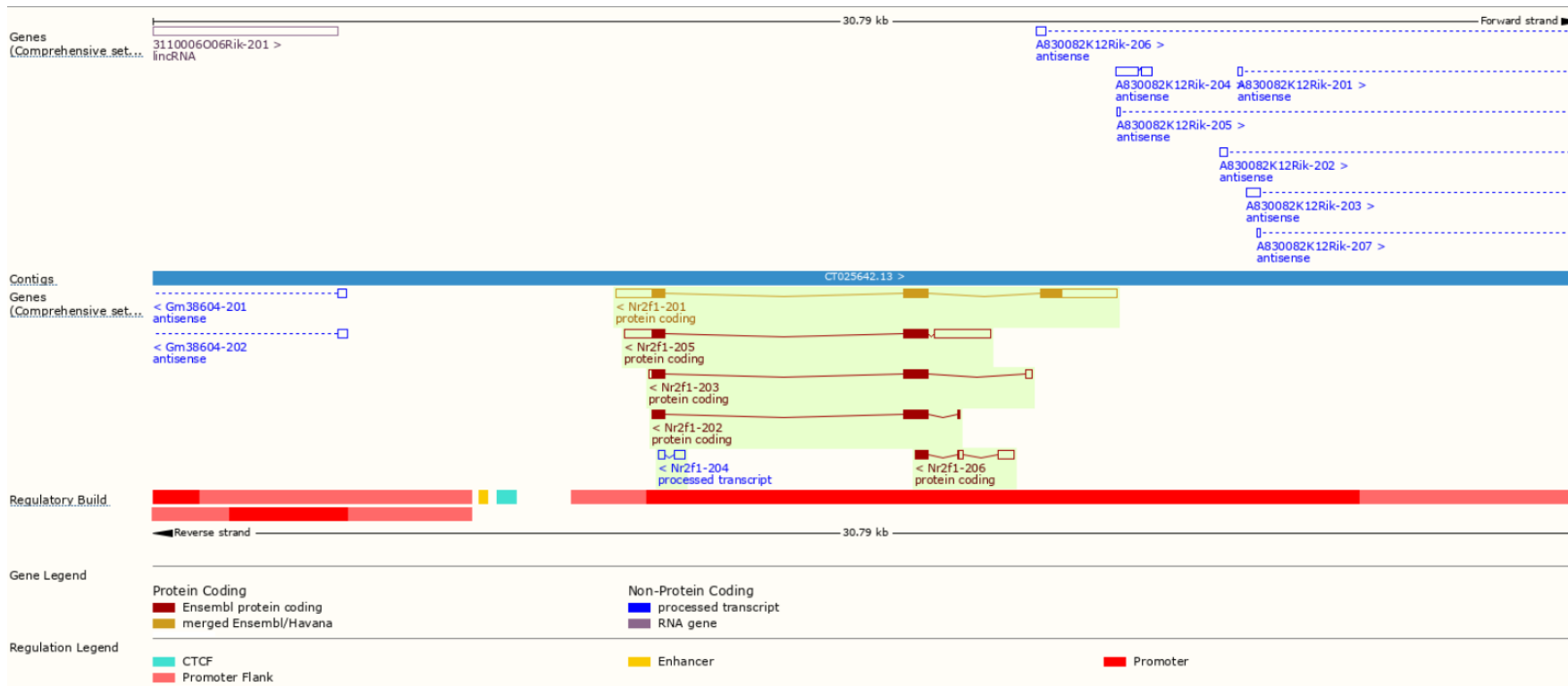


Location: Chromosome 13: 78,188,973-78,199,757

Chr. 13



Gene: Nr2f1 ENSMUSG00000069171

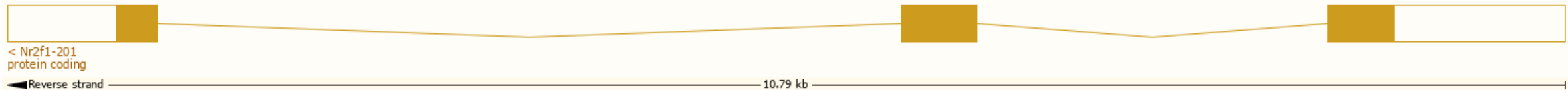


Nr2f1 mRNAs and proteins

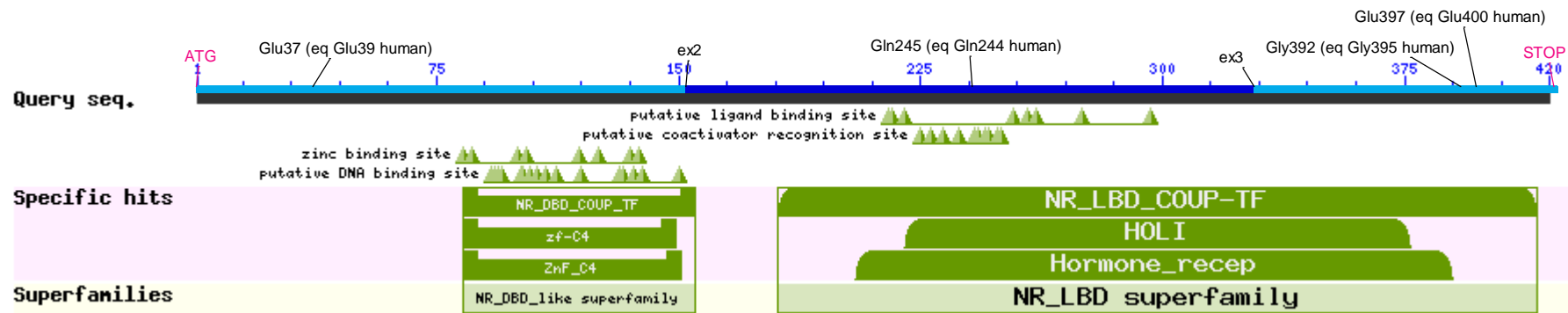


Name	Transcript ID	bp	Protein	Biotype	CCDS	UniProt Match	Flags
Nr2f1-201	ENSMUST00000091458.12	3207	420aa	Protein coding	CCDS26658	Q32NY6	TSL:1GENCODE basicAPPRIS P1
Nr2f1-202	ENSMUST00000125176.2	841	273aa	Protein coding	CCDS84040	F7CII1	TSL:5GENCODE basic
Nr2f1-203	ENSMUST00000127137.2	991	263aa	Protein coding	CCDS84039	B8JJI9	TSL:5GENCODE basic
Nr2f1-204	ENSMUST00000135284.1	368	No protein	Processed transcript	-	-	TSL:3
Nr2f1-205	ENSMUST00000150498.7	2588	263aa	Protein coding	CCDS84039	B8JJI9	TSL:1GENCODE basic
Nr2f1-206	ENSMUST00000224798.1	721	95aa	Protein coding	-	A0A286YD96	CDS 3' incomplete

Transcript: Nr2f1-201 ENSMUST00000091458.12



Positions of the asked mutation in the Nr2f1 protein



Alignment between human and mouse sequence

Query=human (NM_005654.6) vs Subject=Mouse (Nr2f1-201 ENSMUST0000091458.12)

Amino acid alignment

Glu39

Score	Expect	Method	Identities	Positives	Gaps
867 bits(2239)	0.0	Compositional matrix adjust.	419/423(99%)	420/423(99%)	3/423(0%)
Query 1	MAMVVSSWRDPQDDVAGGNPGGNPAAQAARGGGGAGFQQQQAGSGAPHTPQTPGQPGA	60			
Sbjct 1	MAMVVSSWRDPQDDVAGGNPGGNPAAQAARGGGG---HQQQAGSGAPHTPQTPGQPGA	57			
Query 61	PATPGTAGDKGQGGPPGSGSQQHIECVVCGDKSSGKHYGQFTCEGCKSFVKRSVRRNLTY	120			
Sbjct 58	PATPGTAGDKGQGGPPGSGSQQHIECVVCGDKSSGKHYGQFTCEGCKSFVKRSVRRNLTY	117			
Query 121	TCRANRNCPIQHHRNQCQYCRLLKCKLVGMREAVQGRMPPTQPNPGQYALTNGDPLN	180			
Sbjct 118	TCRANRNCPIQHHRNQCQYCRLLKCKLVGMREAVQGRMPPTQPNPGQYALTNGDPLN	177			
Query 181	GHCYLSGYISLLRAEYPYPTSTRYGSQCMQPNMIGIENICELARLLFSAVEWARNIPFF	240			
Sbjct 178	GHCYLSGYISLLRAEYPYPTSTRYGSQCMQPNMIGIENICELARLLFSAVEWARNIPFF	237			
Query 241	PDIQITDQVSLRLRLTWSELFVLNAAQCSMPLHVAPELLAAAGLHASPMSADRVAFMDHIR	300			
Sbjct 238	PDIQITDQVSLRLRLTWSELFVLNAAQCSMPLHVAPELLAAAGLHASPMSADRVAFMDHIR	297			
Query 301	IFQEQVEKALKALHVDSAEYSCLKAIVLFTSDACGLSDAAHIESLQEKSSQCALEEYVRSQY	360			
Sbjct 298	IFQEQVEKALKALHVDSAEYSCLKAIVLFTSDACGLSDAAHIESLQEKSSQCALEEYVRSQY	357			
Query 361	PNQPSRFKGLLLRPLSLRTVSSSVIEQLFFVRLVGGTPEFLIRDMLLSGSSFNWPYMSI	420			
Sbjct 358	PNQPSRFKGLLLRPLSLRTVSSSVIEQLFFVRLVGGTPEFLIRDMLLSGSSFNWPYMSI	417			
Query 421	QCS 423				
Sbjct 418	QCS 420				

Coding sequence alignment

Score	Expect	Identities	Gaps	Strand
1958 bits(1060)	0.0	1201/1269(95%)	9/1269(0%)	Plus/Plus
Query 1	ATGGCAATGGTAGTTAGCAGCTGGCGAGATCCGCAGGACGACGTggccggggggcaacccc	60		
Sbjct 1	ATGGCAATGGTAGTTAGCAGCTGGCGAGATCCGCAGGACGACGTGCCCGGGGGCAACCCC	60		
Query 61	ggggcccccaacccccgagcggcagggcggccggcggggggggggggggggggggggag	120		
Sbjct 61	GGCGCCCCCAACCCCGCAGCGCAGCGACCCCGCGCGCGCGCGCGCGG-----AG	111		
Query 121	caagcagcagggggctggggcgccgcgcaacagcggcagACCCCGGGCCAGCCCGGAGCG	180		
Sbjct 112	CAGCAGCAGGGGGCTCCGGCGCGCCGCACACCGCCGAGACCCCGGGCCAGCCCGGAGCG	171		
Query 181	CCCCCACCACCCCGCAGCGCGGGGACAAAGGGCCAGGGCCCGCCCGGTTCCGGCCAGAGC	240		
Sbjct 172	CCCCCACCACCCCGCAGCGAGGGGACAAAGGGCCAGGGCCCGCCCGGTTCCAGGCCAGAGC	231		
Query 241	CAGCAGCACATCGAGTGCCTGCTGCGGGGACAAAGTGCAGCGGCAAGCACTACGGCCAA	300		
Sbjct 232	CAGCAGCACATCGAGTGCCTGCTGCGGGGACAAAGTGCAGCGGCAAGCACTACGGCCAA	291		
Query 301	TTCACCTGCGAGGGCTGCAAAAAGTTTCTTCAAGAGGAGCGTCCGCAGGAACCTTAACCTAC	360		
Sbjct 292	TTCACCTGCGAGGGCTGCAAAAAGTTTCTTCAAGAGGAGCGTCCGCAGGAACCTTAACCTAC	351		
Query 361	ACATGCCGTGCCAACAGGAACTGTCCCATCGACCAGCACCCCGCAACCACTGCCAATAC	420		
Sbjct 352	ACATGCCGTGCCAACAGGAACTGTCCCATCGACCAGCACCCCGCAACCACTGCCAATAC	411		
Query 421	TGCCGCCTCAAGAAGTGCCTCAAAGTGGGCATGAGGCGGGAAGCGGTTCCAGCAGGAAGA	480		
Sbjct 412	TGCCGCCTCAAGAAGTGCCTCAAAGTGGGCATGAGGCGGGAAGCGGTTCCAGCAGGAAGA	471		
Query 481	ATGCCTCCAACCCAGCCCAATCCAGGCCAGTACGCACTCACCACCGGGGACCCCTCAAC	540		
Sbjct 472	ATGCCTCCAACCCAGCCCAATCCAGGCCAGTACGCACTCACCACCGGGGATCCTCTCAAT	531		
Query 541	GGCCACTGCTACCTGTCCGGCTACATCTCGCTGCTGCTGCGCGCCGAGCCCTACCCACG	600		
Sbjct 532	GGCCACTGCTACCTGTCTGGCTACATTTCTCTGCTGCTGCGCGCAGAGCCCTACCCACG	591		
Query 601	TCGCGCTACGGCAGCCAGTGCATGCAAGCCCAACAACATATGGGCATCGAGAACTCTGC	660		
Sbjct 592	TCGCGTTATGGCAGCCAGTGCATGCAAGCCCAACAACATATGGGCATCGAGAACTCTGC	651		
Query 661	GAGCTGGCCCGCGCCTGCTCTTCAGCGCCGTGAGTGGGCCCGCAACATCCCTTCTTC	720		
Sbjct 652	GAGCTGGCAGCCCGCCTCTCTTCAGCGCCGTGAGTGGGCCCGCAACATCCCTTCTTC	711		
Query 721	CCGGATCTGCAGATCACCAGCCAGGTGTCCCTGCTACGCCCTCACCTGGAGCGAGCTGTT	780		
Sbjct 712	CCGGATCTGCAGATCACCAGCCAGGTGTCTCTGCTGCGCCTCACCTGGAGCGAGCTGTT	771		
Query 781	GTGCTCAACCGCGCCAGTGTCTATGCGCCTGCACGTGGCGCCGTTGCTGGCCCGCCGC	840		
Sbjct 772	GTGCTCAACCGCGCCAGTGTCTATGCGCCTGCACGTGGCGCCGTTGCTGGCCCGCAGCC	831		
Query 841	GGCCTGCATGCTTCGCCCATGTCTGCCGACCCGCTGCTGGCCCTTCATGGACCACATCCGC	900		
Sbjct 832	GGCCTGCACGCTTCGCCCATGTCCGCGGACCCGCTGCTGGCCCTTCATGGACCACATCCGC	891		
Query 901	ATCTTCCAGGACAGGTGGAGAAGCTCAAGGCCTACACGTCGACTCAGCCGAGTACAGC	960		
Sbjct 892	ATCTTCCAGGACAGGTGGAGAAGCTCAAGGCCTGCACGTCGACTCAGCCGAGTACAGC	951		
Query 961	TGCCTCAAAGCCATCGTGTCTTACGCTCAGACGCCTGTGGCCTGTCCGATGCGGCCAC	1020		
Sbjct 952	TGCCTCAAAGCCATCGTGTCTTACGCTCAGATGCTTGTGGCCTGTCCGATGCTGCCAC	1011		
Query 1021	ATCGAGAGCCTGCAGGAGAAGTGCAGTGCAGCTGGAGGAGTACGTGAGGAGCCAGTAC	1080		
Sbjct 1012	ATCGAAAGCCTGCAGGAGAATCACAGTGTCCCTGGAGGAGTATGTGAGAAGCCAGTAC	1071		
Query 1081	CCCAACCAGCCAGCCGTTTGGCAAAGTGTCTGCTGCGACTGCCTCGCTGCGCACCCGTTG	1140		
Sbjct 1072	CCCAACCAGCCAGCCGTTTGGCAAAGTGTCTGCTGCGATTGCGCTCTCTTCGCACAGT	1131		
Query 1141	TCCTCCTCCGTCAATCGAGCAGCTCTCTTCTGCTCCGTTTGGTAGGTAAAACCCCATGAA	1200		
Sbjct 1132	TCCTCCTCTGTCATCGAGCAACTCTTCTGCTACGTTTGGTAGGTAAAACCTCCCATGAA	1191		
Query 1201	ACTCTCATCCGAGATATGTTACTGCTGGAGCAGCTTCAACTGGCCCTTACATGTCCATC	1260		
Sbjct 1192	ACTCTCATCCGAGATATGTTACTGCTCAGGAGCAGCTTCAACTGGCCCTTACATGTCCATC	1251		

Gln244, Gly395; Glu400 are located in very conserved region (aa or nucleotide sequence)
 Glu39* is located in a less conserved region



Selection of the best sgRNA for introduction of Gly395Ala and Glu400Ter

<http://crispor.tefor.net/crispor.py?batchId=9fLgq8zPJacUC9vYfmvW>

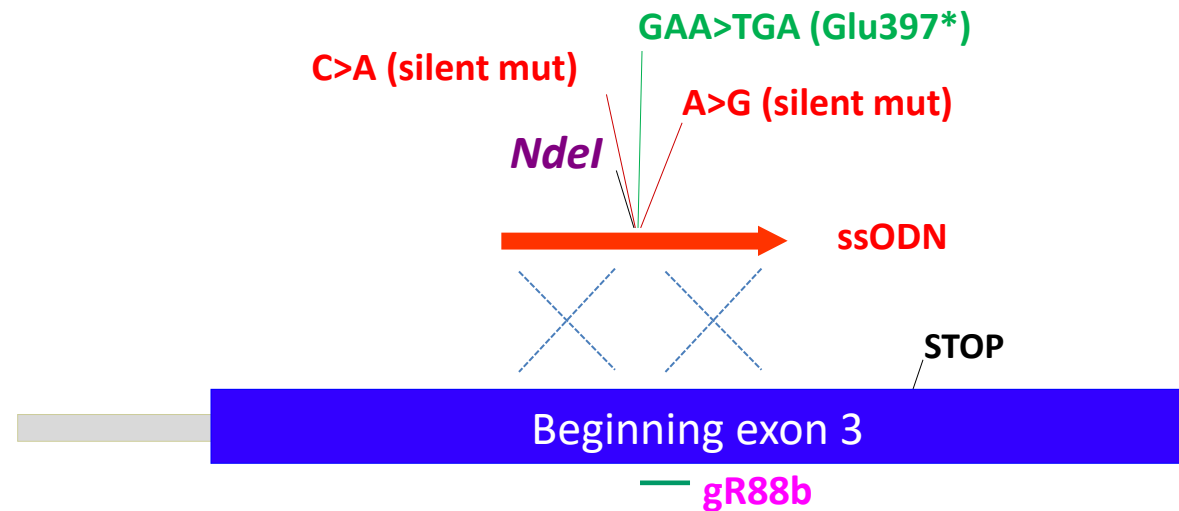
Guide Sequence + PAM + Restriction Enzymes ⓘ + Variants ⓘ <input type="checkbox"/> Only G- <input type="checkbox"/> Only GG- <input type="checkbox"/> Only A- ⓘ	MIT Specificity Score ⓘ	CFD Spec. score ⓘ	Predicted Efficiency ⓘ Show all scores Doench '16 Mor.-Mateos		Outcome Out-of-Frame Lindel		Off-targets for 0-1-2-3-4 mismatches + next to PAM ⓘ	Genome Browser links to matches sorted by CFD off-target score ⓘ <input type="checkbox"/> exons only <input type="checkbox"/> chr13 only
GCAACTCTTCTTCGTACGTT TGG ⚠ Inefficient Enzymes: <i>MaeII</i> , <i>PspI</i> Cloning / PCR primers	97	97	41	51	63	75	0-0-0-1-33 0-0-0-0-0 34 off-targets	4:intergenic:Lmo4-Hs2st1 4:intergenic:Gm12375-Gm12378 4:intergenic:Gm22459-Gm25041 show all...
ATCTCGGATGAGAGTTTCGA TGG Enzymes: <i>TaqI</i> Cloning / PCR primers	91	94	53	38	62	75	0-0-1-2-46 0-0-0-0-0 49 off-targets	2:exon:Nr2f2 4:intergenic:Gm22782-Rpl10l 4:intergenic:Avpr1a-mmu-mir-6412 show all...
CTCTTCTTCGTACGTTTGGT AGG Cloning / PCR primers	88	87	58	20	68	78	0-0-1-7-91 0-0-0-0-0 99 off-targets	4:intron:Slco3a1 4:intron:Axdnd1 4:intergenic:Gm8526-Edil3 show all...
TCTCGGATGAGAGTTTCGAT GGG Enzymes: <i>TaqI</i> Cloning / PCR primers	88 b	94	69	46	63	66	0-0-1-8-60 0-0-0-0-0 69 off-targets	4:intergenic:Gm5382-Gm14867 2:exon:Nr2f2 4:intergenic:9530052E02Rik-Gm15418 show all...

Haeussler, M. et al. Evaluation of off-target and on-target scoring algorithms and integration into the guide RNA selection tool CRISPOR. *Genome Biol.* **17**, 148. doi:10.1186/s13059-016-1012-2

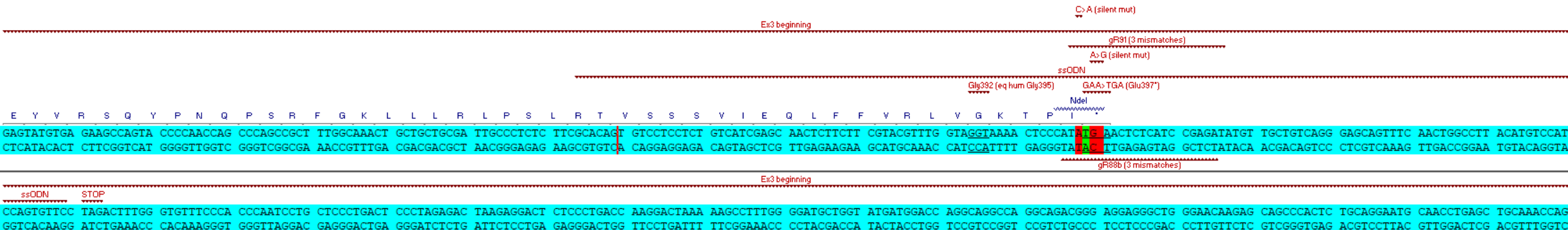
Strategy validated: introduction of Gln397* (corresponding to Glu400* in human)

Validated by the client

Approach proposed: Homology directed repair (HDR) via CRISPR/Cas9



Sequence of the allele after HDR



Strategy validated: introduction of Gln397* (corresponding to Glu400* in human)



■ PROS

- Two silent mutations (one in the STOP codon) will be added to the asked G>T mutation in order to avoid a recut after HDR. The 3 mutations will introduce a new NdeI diagnostic restriction site that will easy genotyping.

■ CONS

- Frequency of HDR is not predictable and can be low

Line Kus7772-1-PM

F1 genotype

Sequence with PCR F1-R1

Primers in orange

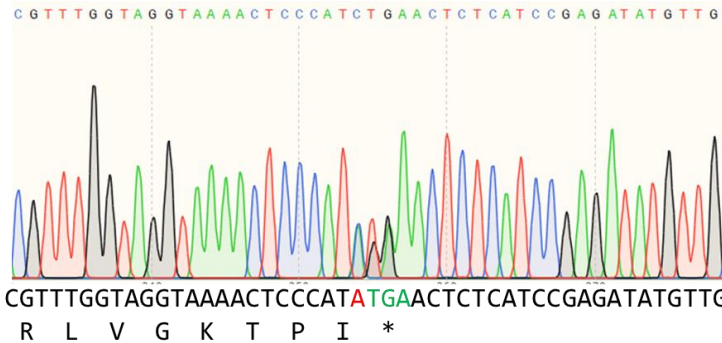
Asked mutation in green

Silent mutation in red

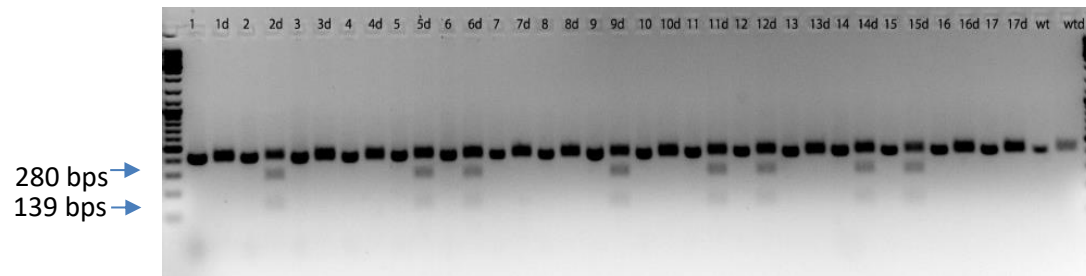
NdeI site in italic and underlined

TG**TATGCCTTTCCATCTCACTCACCT**TAACAATCTGTCCCCTTTCCCTGTCTCTCCCTCCTGTGGCTGCTCTGGCAGATGCTTGTGGCCTGTCGGATGCTGCCACATCGAAAGCCTG
 CAGGAGAAATCACAGTGTGCCCTGGAGGAGTATGTGAGAAGCCAGTACCCCAACCAGCCCAGCCGCTTTGGCAAAGTCTGCTGCTGCGATTGCCCTCTCTTCGCACAGTGTCCCTC
 CTCTGTCATCGAGCAACTCTTCTTCGTACGTTTGGTAGGTAAACTCCCAT**ATG**AACTCTCATCCGAGATATGTTGCTGTCAGGGAGCAGTTTCAACTGGCCTTACATGTCCATCCA
 GTGTTCTAGACTTTGGGTGTTTCCCACCCAATCCTGCTCCCTGACT**CCCTAGAGACTAAGAGGACTCTCCC**

Sanger sequencing (F1 #2)



F1 genotyping



The line **Kus7772-1-PM** was established. Four heterozygous males and 4 heterozygous females are available. Heterozygous pups with the premature STOP codon are viable at the heterozygous stage.

International nomenclature (according to MGI <http://www.informatics.jax.org/mgihome/nomen/gene.shtml>) : **Nr2f1^{em1lcs}**

MGI allele ID: 8253350

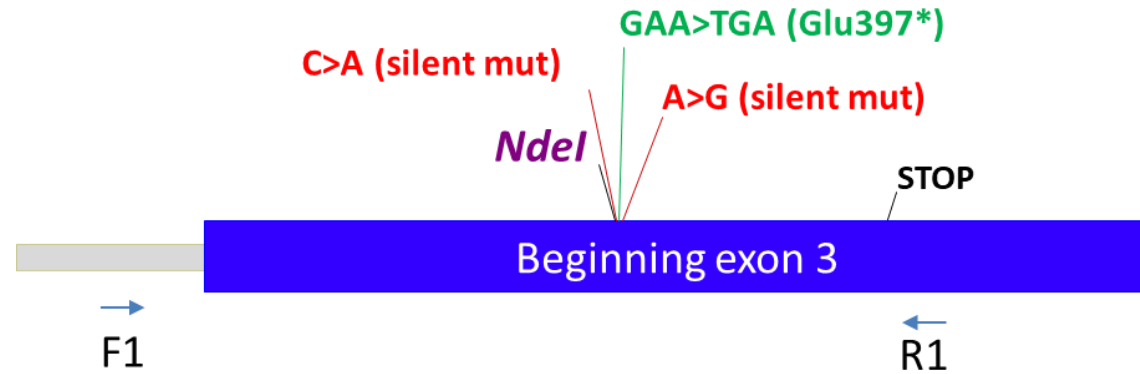
Genotyping protocol



GENOTYPING INSTRUCTIONS



Genotyping scheme



PCR genotyping primers and sequence

Primer ref.	Sequence	Amplification product size WT	Sizes observed if Ndel restriction present (associated with asked PM)
F1	TGTATGCCTTTCCATCTCACTCACC	419 bps	139 +280 bps
R1	GGGAGAGTCCTCTTAGTCTCTAGGG		

Sequence with PCR F1-R1

Primers in orange

Asked mutation in green

Silent mutation in red

Ndel site in italic and underlined

TGTATGCCTTTCCATCTCACTCACC TAACAATCTGTCCCCTTTCCCTGTCTCTCCCTCCTGTGGCTGCTCTGGCAGATGCTTGTGGCCTGTCGGATGCTGCCACATCGAAAGCCTG
 CAGGAGAAATCACAGTGTGCCCTGGAGGAGTATGTGAGAAGCCAGTACCCCAACCAGCCCAGCCGCTTTGGCAAAGTCTGCTGCTGCGATTGCCCTCTCTTCGCACAGTGTCTC
 CTCTGTCATCGAGCAACTCTTCTTCGTACGTTTGGTAGGTA AA ACTCCCATATGAACTCTCATCCGAGATATGTTGCTGTCAGGGAGCAGTTTCAACTGGCCTTACATGTCCATCCA
 GTGTTCTAGACTTTGGGTGTTTCCCACCCAATCCTGCTCCCTGACTCCCTAGAGACTAAGAGGACTCTCCC

PCR Protocol

This section describes the composition of the mix and the cycling conditions used for genotyping F0 and F1 genotyping.

Reagents:	Volume (per sample):
- Phusion HS (Thermo Scientific) 5X Buffer	4 μ l
- 10mM dNTP	0.4 μ l
- 5' primer (100 μ M)	0.1 μ l
- 3' primer (100 μ M)	0.1 μ l
- DNA (lysate 1/10)	2 μ l
- Phusion Hot Start II	0.2 μ l
- Sterile H2O	up to 20 μ l

Cycling conditions

Temp	Time	#Cycles
96°C	5min	1
96°C	8s	
62°C	10s	30
68°C	45s	
68°C	5min	1
12°C	5min	1

Digestion protocol	Volume / sample
PCR product	10 μ l
Buffer 10X	2 μ l
Restriction enzyme	0.2 μ l
H2O	7.8 μ l

This reaction is incubated 15 mins at 37°C then loaded on a 3% agarose. The 10 μ l left over PCR reaction serves as negative control



REPORT REDACTION & VALIDATION

Protocol finalized on 2021/ 08 /24

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