



R105Q KI in Orc1 by CRISPR-Cas9 approach and genotyping protocol

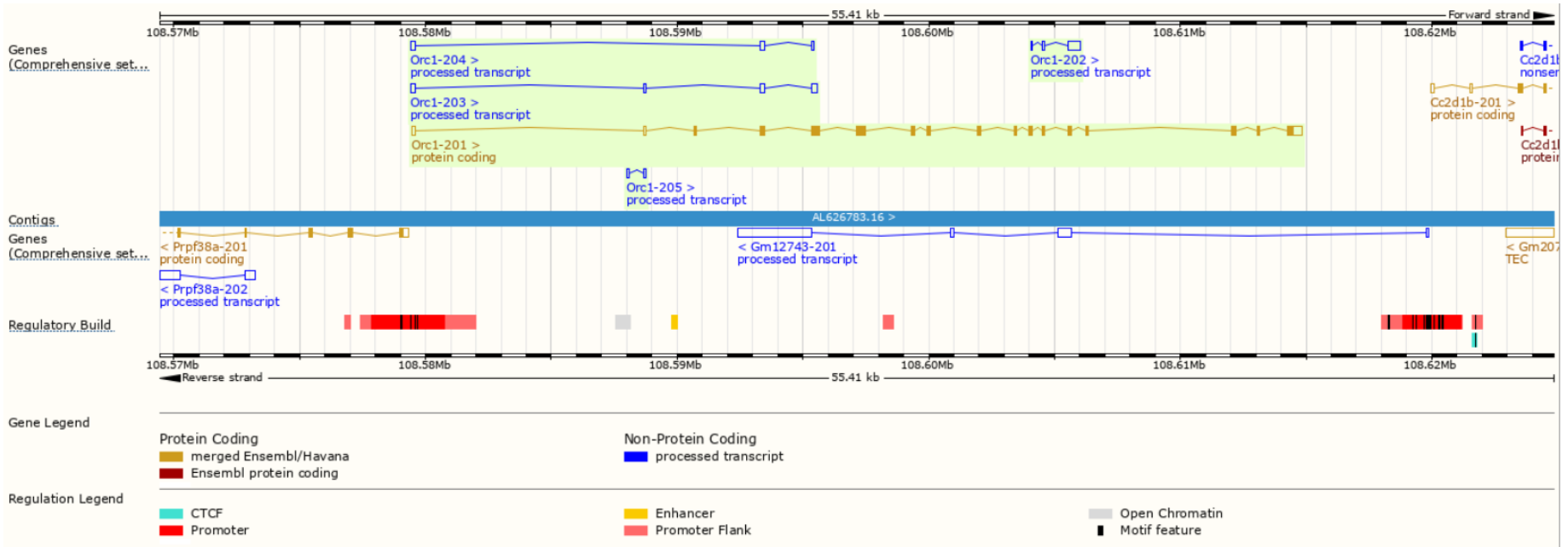
Kus6784 / IM6784

Contact: mutagenesis@igbmc.fr

Orc1 Mouse Genomic locus



Gene: Orc1 ENSMUSG00000028587



Chromosome 4: 108,579,423-108,614,833



Orc1 mRNAs and proteins



Name	Transcript ID	bp	Protein	Biotype	CCDS	UniProt	RefSeq
Orc1-201	ENSMUST00000102744.3	3014	840aa	Protein coding	CCDS18453	Q9Z1N2	NM_011015 NP_035145
Orc1-202	ENSMUST00000129931.1	670	No protein	Processed transcript	-	-	-
Orc1-203	ENSMUST00000130162.1	679	No protein	Processed transcript	-	-	-
Orc1-204	ENSMUST00000139772.7	443	No protein	Processed transcript	-	-	-
Orc1-205	ENSMUST00000143497.1	173	No protein	Processed transcript	-	-	-

421 T T G C T ^{Y*} T G A G T T G ^{K YR} T T C G A A G A T ^{*} G A C T C T G A T ^{SY B} C C T C C T C T A A G A A A C ^Y C G T G C T C G A G T A C A G 480
202 T T G C T T G A G T T G T T C G A A G A T G A C T C T G A T C C T C C T C T A A G A A A C G T G C T C G A G T A C A G 261
68 -L--L--E--L--F--E--D--D--S--D--P--P--P--K--K--R--A--R--V--Q-- 87

481 T G G T T T G T C ^Y C G A T T C T G T G A A G T C C C T G ^{YR} C C T G T A A A C G G C A T T T G T T G G G C C G G A A G C C T 540
262 T G G T T T G T C C G A T T C T G T G A A G T C C C T G C C T G T A A A C G G C A T T T G T T G G G C C G G A A G C C T 321
88 -W--F--V--R--F--C--E--V--P--A--C--K--R--H--L--L--G--R--K--P-- 107

541 ^{*} G G T G C A C A G G A A A T A T T C T G G T A T G A T T A C C C G G C C T G T G A C A G C A A C A T T A A T G C G G A G 600
322 G G T G C A C A G G A A A T A T T C T G G T A T G A T T A C C C G G C C T G T G A C A G C A A C A T T A A T G C G G A G 381
108 -G--A--Q--E--I--F--W--Y--D--Y--P--A--C--D--S--N--I--N--A--E-- 127

Human ORC1 (Query) vs Mouse Orc1 (Subject)

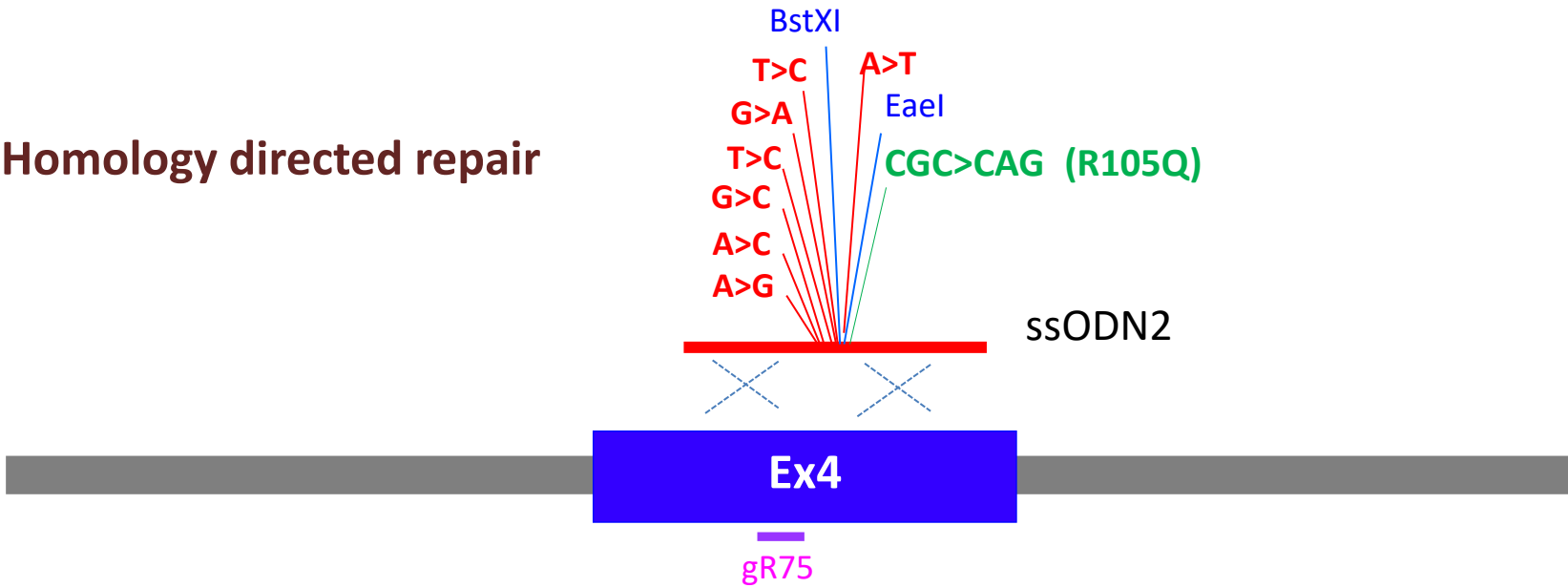


Query	1	MAHYPTRLKTRKTYSWVGRPLLDRLKHYQTYREMCVKTEGCSSTEIHIQIGQFVLIQEGDDD	60
Sbjct	1	M Y TR KTR+T+SWVGRPL +RK Q YRE+C+K S EIHI++GQFVLI+G+D+ MPSYLTRQKTRQTFSWVGRPLPNRKQFQQMYREICMKINDGS-EIHIKVGQFVLIQGEDN	59
Query	61	ENPYVAKLLELFEDDSDPPPKKRARVQWVFRFCEVPACKRHLLGRKPGAQEIFWYDYPAC	120
Sbjct	60	+ PYVAKL+ELF++ ++ PPKK ARVQWVFRF E+P KRHLLGR P AQEIFWYD KKPYVAKLIELFQNGAEVPPKKCARVQWVFRFLEIPVSKRHLLGRSPPAQEIFWYDCSDW	119
Query	121	DSNINAETIIGLVRVIPLAPKDVVPTNLKNEKTLFVKLSWNEKKFRPLSSELFAELNKPQ	180
Sbjct	120	D+ IN ETIIG V+V+ LAP++V+P + K+E+TLFVKLSWN+K F PL E+ A L + Q DNKINVTIIGPVQVVALAPEEVI PV DQKSEETL FVKLSWNKKDFAPLPPEVLAAL-REQ	178
Query	181	ESAAKQKQFVRAKSASAESPSWTPAEHVAKRIESRHSASKSRQTPHPLTPRARKRELELG	240
Sbjct	179	E + + QKP++AK K+ +SP+ E K I+S HS SK QTP + + P A+K LEL EDSPEWQKPLKAKIKNVKSPARNTTEQEVKGIKSNHSTSKFHQTPANIVIPNAKKSLELD	238
Query	241	NLG-----NPQMSQQTSCASLDSPGRIKRVAFSEITSPSKRSQPKLQTLSP--ALKAP	293
Sbjct	239	LG N + S+++SC SLD KR+ AFSE TSP K+ P+K + + P AL+ GLGFTRPKNTRWSKSSCDSDLDYQKTSKRRAAFSETTSPPKK--PNKPREIKPSSALETR	296
Query	294	EKTRETGLSYTEDDKKASPEHRIILRTRIAASKTIDIREERTLTPISGGQRSSVVPSVIL	353
Sbjct	297	K +T + + ++LR R A T + + TL+PI G RSSVPS L VKNGQT-----QPFCAKSSVVLRRARNPAMTTTKLGV DNTLSPIRNGLRSSVVPSGGL	348
Query	354	KPENIKRDAKEAKAQNEATSTPHRIRKSSVLTMRIRQQRLRFLGNSKSDQEEKEILPA	413
Sbjct	349	P I+ R AKE + E T R+ RKSS+LT+ RIRQQ L DQE TPVYIR-RKAKEQETHKEPIRT-SRVHRKSSLLTLKRIRQQCLLDGDDRDQE-----E	400
Query	414	AEISDSSSDEEEASTPPLPRRAPRTVSRNLRSSLKSSLHTLTKVPKSLKPRTPRCAAPQ	473
Sbjct	401	E DS S+EE+ LP R SR + T +K P+K+ KPRT P A PQ EESVDSESEEEDEFISSLPTRNLSLQGSR-----TRQTPSKSPQKNPKPRTPHRATPQ	452
Query	474	IRSRSLAAQEPASVLEEARLRLHVS AVPESLPCREQEFQDIYNFVESKLLDHTGGCMYIS	533
Sbjct	453	IR R+LA QEPAS LEEARLRLHVS AVP+SLPCREQEFQDIY+ FVESKLLD TGGCMYIS IRDRNLAVQEPASALEEARLRLHVS AVPDSLPCREQEFQDIYSFVESKLLDGTGGCMYIS	512
Query	534	GVPGTGKTATVHEVIRCLQQAQANDVPPFQYIEVNGMKLTEPHQVYVQILQKLTGQKAT	593
Sbjct	513	GVPGTGKTATVHEVIRCLQQA+ +DVPPFQY+EVNGMKLTEPHQVYVQIL+KLTGQKAT GVPGTGKTATVHEVIRCLQQAETDDVPPFQYVEVNGMKLTEPHQVYVQILKLTGQKAT	572
Query	594	ANHAAELLAKQFCTRGSPQETTLLVDEL DLLWTHKQDIMYNLFDWPTHKEARLVVLAIA	653
Sbjct	573	ANHAAELLAKQFC +GS +ETTLLVDEL DLLWTHKQD+MYNLFDPWPTH A L+VL IA ANHAAELLAKQFCGQGSQKETTLLVDEL DLLWTHKQDVMYNLFDWPTHGAHLIVLTIA	632
Query	654	NTMDLPERIMMNRVSSRLGLTRMCFQPYTYSQLQQILRSRLKHLKAFEDDAIQLVARKVA	713
Sbjct	633	NTMDLPERIMMNRVSSRLGLTRM FQPY++SQL+QIL SRL++L+AFEDDAIQLVARKVA NTMDLPERIMMNRVSSRLGLTRMSFQPYSHSQLKQILVSRRLNRAFEDDAIQLVARKVA	692
Query	714	ALSGDARRCLDICRRATEICEFSQQKPDSPGLVTIAHSMEAVDEMFSSSYITAIKNSSVL	773
Sbjct	693	ALSGDARRCLDICRRATEICE S + DS LVT+AH MEA+DEMFSSSYITAIKNSSV+ ALSGDARRCLDICRRATEICELSHLRGDSLSLVTVAHLMEAIDEMFSSSYITAIKNSSV	752
Query	774	EQSFLRAILAEFRRSGLLEATFQQIYSQHVALCRMEGLPYPTMSETMAVCSHLSGCRLLL	833
Sbjct	753	EQSFLRAI+AEFRRSGLLEATFQQIYSQHVALCRMEGLPYPTMSETMAVCS LGSCRLLL EQSFLRAIIAEFRRSGLLEATFQQIYSQHVALCRMEGLPYPTMSETMAVCSRLGSCRLLL	812

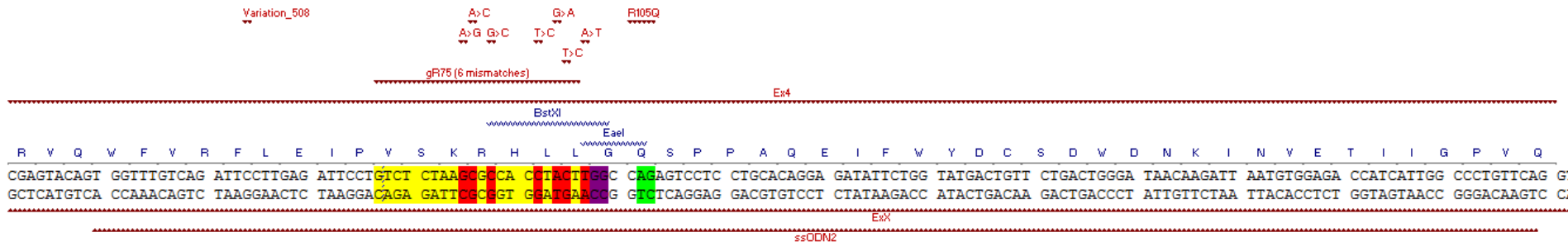
Proposal: introduction of the R105Q PM in Orc1



Homology directed repair



Expected allele



New diagnostic restriction sites : BstXI, EaeI (best choice)

Line Kus6784-38-PM



F1 genotype

Sequence with PCR F1-R1

Oligo F and R in orange

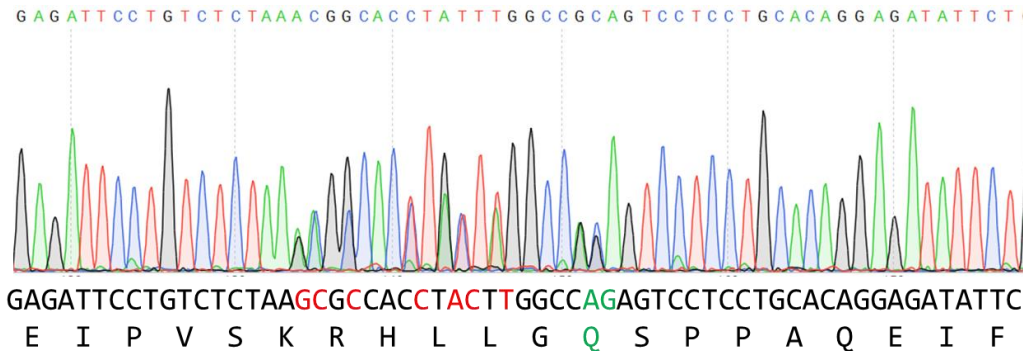
Silent mutations

Asked PMs in green

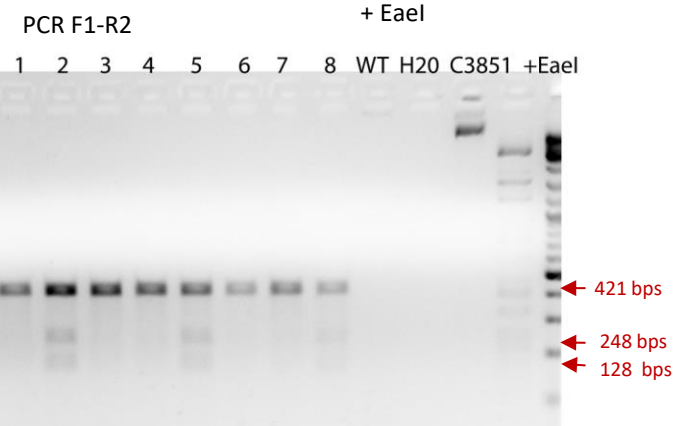
Diagnostic restriction site in italic

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 GAGCTGAAGTTCCTCCCAAGAAATGTGCACGAGTACAGTGGTTTGTGAGATTCTTGAGATTCTGTCTCTAAGCGCCACCTACTT
 GGCCAGAGTCCTCCTGCACAGGAGATATTCTGGTATGACTGTTCTGACTGGGATAACAAGATTAATGTGGAGACCATCATTGGCCC
 TGTTTCAGGTTTCGTGCCTTAAATCCTGCTTCTATCTTTAGCACCCACACTGCATGGCTTCAGAAATATTACTAGTTCTAATTTGAATTC
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Sanger sequencing

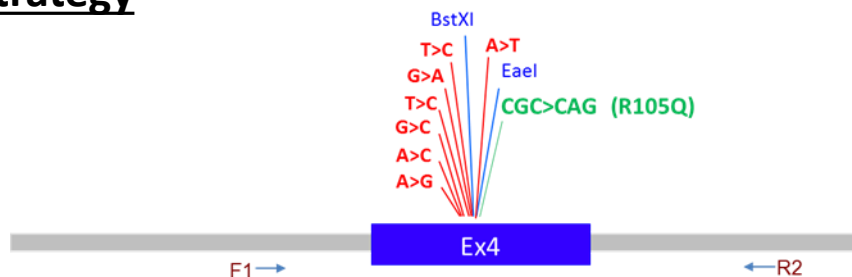


F1 genotyping



FO	Nb F1 born	F1 heterozygote	
		M	F
Kus7015-25	46	6	6

PCR genotyping strategy



Primer ref.	Sequence	Amplification product size WT	Expected size if BstUI diagnostic restriction is present
F1	GTTGTGTAGTGTTTGACACGTAGGC	421 bps	248 bps + 173 bps
R2	AGCCAGAGCATTATGTTCTATGCT		

Sequence of the PCR product (with mutations introduced)

GTTGTGTAGTGTTTGACACGTAGGC AATATTCAGTGCTGTATTGTTTGCCTTACTATTTAATCTTCCC GCCCTCCCCAATCCTCCT
 AGGAGCTGAAGTTCCTCCCAAGAAATGTGCACGAGTACAGTGGTTTGT CAGATTCCTTGAGATTCCTGTCTCTAAGCGCCAC
 C TACTTGGCCAGAGTCCTCCTGCACAGGAGATATTCTGGTATGACTGTTCTGACTGGGATAACAAGATTAATGTGGAGACCAT
 CATTGGCCCTGTT CAGGTTCTGCCTTAAATCCTGCTTCTATCTTTAGCACCCACACTGCATGGCTTCAGAAATATTACTAGTTC
 TAATTTGAATTCTGTTCTAATTATCTGTTTGGCTGCTTGCCCCAGCACACAGATACTGACAGCATAGAACATAATGCTCTGGGCT

PCR Protocol

This section describes the composition of the mix and the cycling conditions used for genotyping F0.

Reagents:	Volume (per sample):
- Phusion HS (Thermo Scientific) 5X Buffer	4 μ l
- 10mM dNTP	0,4 μ l
- 5' primer (100 μ M)	0.1 μ l
- 3' primer (100 μ M)	0.1 μ l
- DNA (lysate 1/10)	2 μ l
- Phusion Hot Start II	0,2 μ l
- Sterile H ₂ O	up to 20 μ l

Cycling conditions

Temp	Time	#Cycles
96°C	5min	1
96°C	8s	
62°C	10s	30
68°C	45s	
68°C	5min	1
12°C	5min	1

Digestion protocol	Volume / sample
PCR product	10 μ l
Buffer 10X	2 μ l
Restriction enzyme EaeI	0,2 μ l
H ₂ O	7,8 μ l

This reaction is incubated 15 mins at 37°C then leaded on a 3% agarose. The 10 μ l left over PCR reaction serves as negative control.