



# Introduction of the C395Y point mutation in mouse Cckar (corresponding to C387Y in human) Genotyping protocol

**Kus7497**

**CALL PHENOMIN-FMR LAUREATE PROJECT**

Done by Marie-Christine Birling (PhD)

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## Approach validated





# Introduction of the C395Y point mutation in mouse Cckar (corresponding to C387Y in human) via a CRISPR/Cas9 approach

**Kus7497 / IM7497**

Strategy proposed by Marie-Christine Birling (PhD)

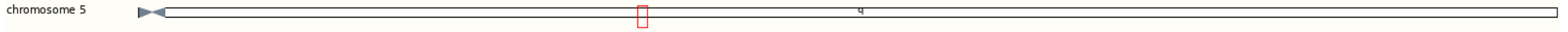
[birlingm@igbmc.fr](mailto:birlingm@igbmc.fr)

18/01/2022

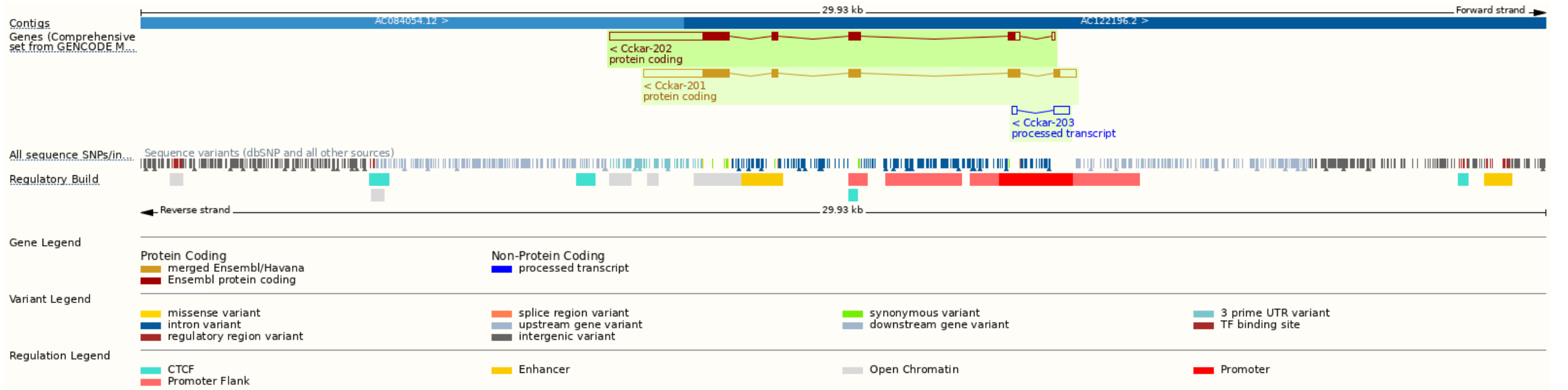
# Cckar Mouse Genomic locus



Location: Chromosome 5: 53,855,118-53,865,047



## Gene: Cckar ENSMUSG0000029193



# Cckar mRNAs and proteins



Transcript ID	Name	bp	Protein	Biotype	CCDS	UniProt Match
<a href="#">ENSMUST00000031093.5</a>	Cckar-201	2930	<a href="#">436aa</a>	Protein coding	<a href="#">CCDS19293</a>	<a href="#">O08786</a>
<a href="#">ENSMUST00000200691.4</a>	Cckar-202	3230	<a href="#">365aa</a>	Protein coding	<a href="#">CCDS84881</a>	<a href="#">Q3TPLO</a>
<a href="#">ENSMUST00000202946.2</a>	Cckar-203	424	No protein	Processed transcript	-	-

**Transcript: ENSMUST00000031093.5 Cckar-201**

# Strategy proposed: Selection of the best sgRNAs



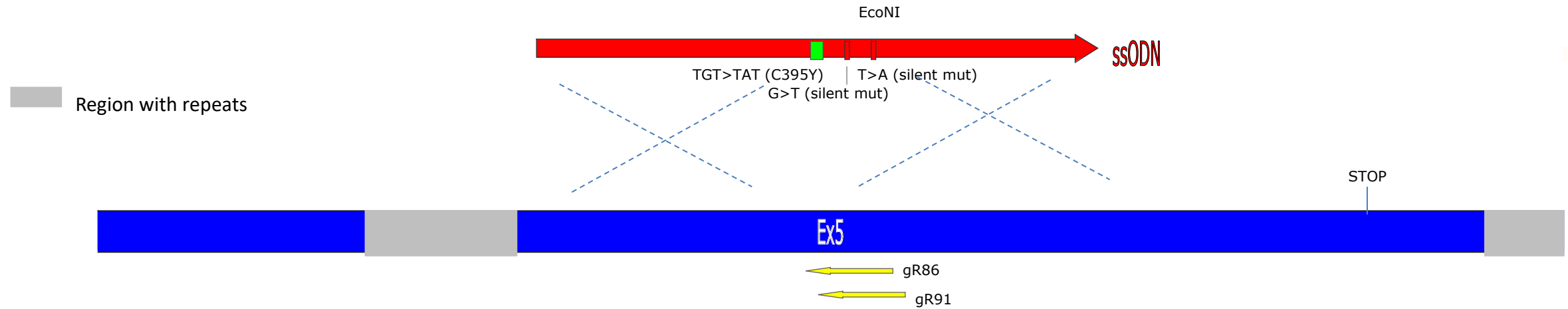
<http://crispor.tefor.net/crispor.py?batchId=5KGHtECzxlCjylPvm1J>

Guide Sequence + PAM + Restriction Enzymes ⓘ + Variants ⓘ <input type="checkbox"/> Only G- <input type="checkbox"/> Only GG- <input type="checkbox"/> Only A- ⓘ	MIT Specificity Score ⓘ	CFD Spec. score ⓘ	Predicted Efficiency ⓘ <small>Show all scores</small> Doench '16 Mor.-Mateos		Outcome Out-of-Frame Lindel		Off-targets for 0-1-2-3-4 mismatches + next to PAM ⓘ	Genome Browser links to matches sorted by CFD off-target score ⓘ <input type="checkbox"/> exons only <input type="checkbox"/> chr5 only
GGGACCAGGATTTCGGGCAAC AGG ..... <b>Cloning / PCR primers</b>	91	95	31	56	72	75	0-0-1-4-59 0-0-0-0-0 64 off-targets	4:intron:Stra6 4:intergenic:Tmem154-Fbxw7 4:intergenic:Mapk4/Gm9925-Ska1 show all...
GACCAGGATTTCGGGCAACAG GGG ..... <b>Cloning / PCR primers</b>	86	94	68	67	68	74	0-0-1-4-72 0-0-0-0-0 77 off-targets	4:intergenic:Gm22011-Mthfr-ps1 4:intron:Snx3 4:intergenic:Mir1971-Gm23652 show all...

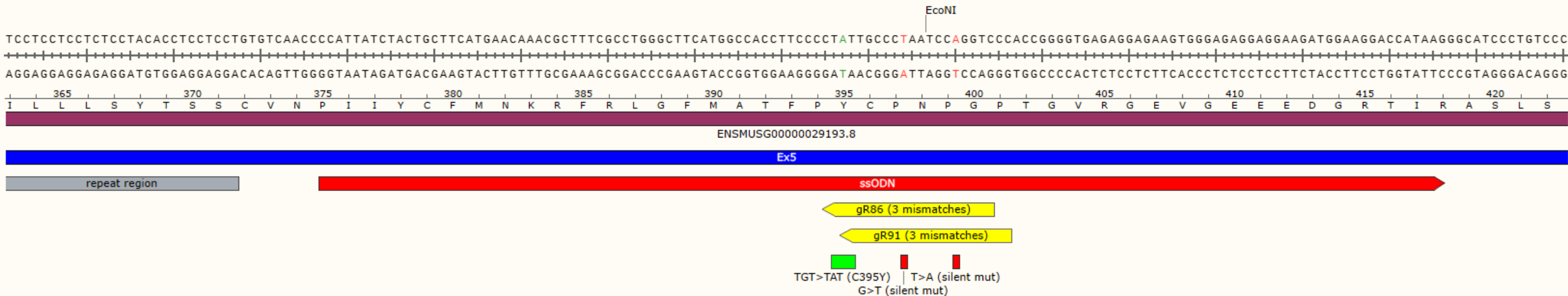
Haeussler, M. et al. Evaluation of off-target and on-target scoring algorithms and integration into the guide RNA selection tool CRISPOR. *Genome Biol.* **17**, 148. doi:10.1186/s13059-016-1012-2

# Strategy proposed: introduction of the Cys395Tyr point mutation via CRISPR/Cas9

Approach proposed: homology directed repair (HDR) after CRISPR directed double strand break in fertilized oocytes



## Sequence detail of the locus after HDR



# Strategy proposed: introduction of the Cys395Tyr point mutation via CRISPR/Cas9

## ■ PROS

- The double strand break obtained with CRISPR guide RNA gR86 or gR91 are ideally located close to the mutation to introduce
- Two additional silent mutations (C>T and T>A) will be introduced in order to avoid a new CRISPR guided double strand break after HDR. Three mismatches will be introduced in total in the donor ssODN.
- The silent mutations will also allow the introduction of a new diagnostic restriction site

## ■ CONS

- Frequency of HDR is not predictable and can be low

# Genotyping protocol



## F1 genotyping

F1 and R1 are in orange

Silent mutation is in red

Asked mutation leading to the C395Y mutation is in green

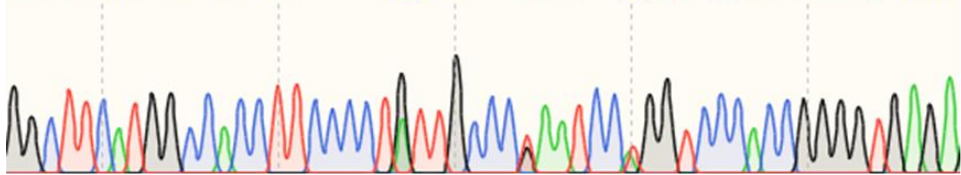
*EcoNI* restriction sites are in italic and underlined

CATATGACACGGTTTCTGCCGAGAAACACCTCTCAGGGACCCCATCTCCTTCATCCTCCTCCTCCTACACCTCCTCCTGTGTCAACCCCATCTACTGCTTCATGAACAAACGCTTTGCGCTGGGCTTCATGGCCACCTTCCCCTATTGCCCTAATCCAGGTCCCACCGGGGTGAGAGGAGAAGTGGGAGAGGAGGAAGATGGAAGGACCATAAGGGCATCCCTGTCCCAGTATTCTACAGCCACATGAGCACCTCTGCCACCCCATGAGCTGTACCTGGTTCGCTGCGGGCAGCAGAAAGGAGGGGACTGGGAGCAAGGAGGAAGAGGAAGTGGGAGGGAAGGAGAAGGAAGACCCATTTCTAACGAGGACTCTCCGATGTCTGCTTTT

## Sanger sequencing

F1#2

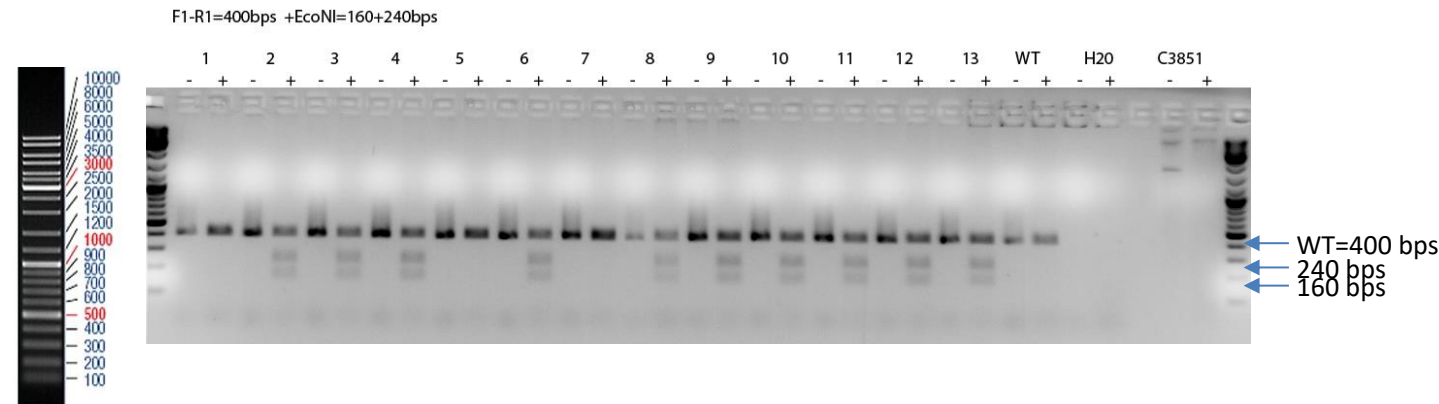
GGCTTCATGGCCACCTTCCCCTGTTGCCCTAATCCGGTCCCACCGGGGTGAGA



GGCTTCATGGCCACCTTCCCCTATTGCCCTAATCCAGGTCCCACCGGGGTGAGA

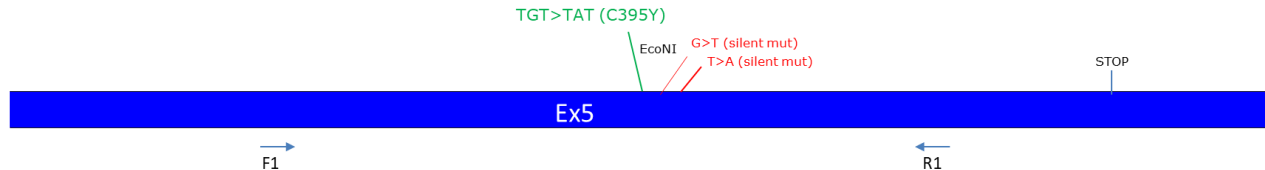
G F M A T F P Y C P N P G P T G V R

## F1 genotyping



The line Kus7497-13-PM is established and cryopreserved

## Genotyping Scheme



## PCR genotyping primers and sequence

Primer ref.	Sequence	Amplification product size WT	Sizes observed if new EcoNI restriction present (associated with asked PM)
F1	CATATGACACGGTTTCTGCCGAGAA	400 bps	240 bps + 160 bps
R1	AAAAGCAGACATCGGAAGAGTCCTC		

**CATATGACACGGTTTCTGCCGAGAA**ACACCTCTCAGGGACCCCATCTCCTTCATCCTCCTCCTCCTACACCTCCTCCTGTGTCAACCCATTATCTACTGCTT  
 CATGAACAAACGCTTTCGCCTGGGCTTCATGGCCACCTTCCCCTATTGCCCT**TAATCCA**AGGTCCCACCGGGGTGAGAGGAGAAGTGGGAGAGGAGGAAGATG  
 GAAGGACCATAAGGGCATCCCTGTCCCGGTATTCTACAGCCACATGAGCACCTCTGCCCCACCCCACTGAGCTGTACCTGGTTTCGCTGCGGGCAGCAGAAA  
 GGAGGGGACTGGGAGCAAGGAGGAAGAGGAAGTGGGAGGGGAAGGAGAAGGAAGACCCATTTCTAACGAG**GACTCTTCCGATGTCTGCTTTT**

F1 and R1 are in orange

Silent mutation is in red

Asked mutation leading to the C395Y mutation is in green

EcoNI restriction sites are in italic and underlined

## PCR Protocol

This section describes the composition of the mix and the cycling conditions used for genotyping F0 and F1 genotyping.

Lysis buffer: 50 µl DNA Extract All Reagents – appliedbiosystems, Ref 4402616 (25 µl Lysis buffer +25 µl stabilising buffer)

### Reagents:

	Volume (per sample):
- Phusion HS (Thermo Scientific) 5X Buffer	4 µl
- 10mM dNTP	0.4 µl
- 5' primer (100 µM)	0.1 µl
- 3' primer (100 µM)	0.1 µl
- DNA (lysate 1/10)	2 µl
- Phusion Hot Start II	0.2 µl
- Sterile H2O	up to 20 µl

### Cycling conditions

Temp	Time	#Cycles
96°C	5 min	1
96°C	8s	30
62°C	10s	
68°C	45s	
68°C	5min	1
12°C	5min	1

### Digestion protocol

### Volume / sample

PCR product	10 µl
Buffer 10X	2 µl
Restriction enzyme	0.2 µl
H2O	7.8 µl

This reaction is incubated 15 mins at 37°C then leaded on a 3% agarose. The 10 µl left over PCR reaction serves as negative control



## REPORT REDACTION

Report performed on 2022/12/19  
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