

EMMA health monitoring procedures

Brief description of housing system, health monitoring programme and health status of the EMMA SPF live colonies

Type of facility

Animals distributed by EMMA are bred in SPF (Specific Pathogen Free) barriered facilities in which all materials are sterilized before entry. Staff entering the barriered areas must shower and change into clean unit clothing. Where appropriate, staff working within the units is also required to wear, gloves, face masks, mob caps and over shoes.

Housing system

Animals are maintained in either flexible film isolators or IVCs (Individually Ventilated Cages) or in conventional cages in barriered areas under positive pressure and are given autoclaved bedding, autoclaved or irradiated food and filtered or chlorinated water. Animals reared in IVCs are cage changed under laminar flow hoods.

Sentinel programme

The health status of each animal room is monitored on a regular basis e.g. 4 times per year when mice are reared in IVCs or monthly when mice are bred in conventional cages in barriered areas. These screening programmes involve exposing sentinel animals to dirty bedding collected from other IVCs within the mouse room. Some colonies e.g. those reared in isolators are sampled directly.

Health report

Before receiving any mice from EMMA you will be sent a recent (< 3 months old) health report prepared in accordance with the FELASA (Federation of European Laboratory Animal Science) recommendations. This health report will give details of the agents tested, the number of animals tested and the analytical methods used.

The following pages provide a sample health report from the EMMA node that distributes the strain you are interested in. Note that this is a **sample** health report and **not a current report**. Current reports will be provided upon request. Additional specific health checks (beyond tests recommended by FELASA) are possible if required by customers for importation but will be charged to the customer. If you require any further information please contact the archiving/distribution centre handling your request.



"ALEXANDER FLEMING"
Biomedical Sciences Research Center

Health Monitoring Report

(Date: 21.07.2020)

Location: Core Breeding Unit
Strain: wt, KO, KI & Tg

Housing: Filter top cages/changing station
Testing laboratories: Idexx Laboratories

Species: Mouse

Latest test date: July 2020

Next Screening: October 2020

	07/2020	METHOD	04/2020	01/2020	11/2019	08/2019	05/2019	02/2019
VIRUSES								
Ectromelia virus			0/2		0/2		0/2	
Hantavirus	0/2	MFI-serology	0/2	0/2	0/2	0/2	0/2	0/2
Lactate dehydrogenase elevating virus					0/2		0/2	
Lymphocytic choriomeningitis virus					0/2		0/2	
Minute virus of mice	0/2	MFI-serology	0/2	0/2	0/2	0/2	0/2	0/2
Mouse adenovirus type 1 (Mad FL)			0/2		0/2		0/2	
Mouse adenovirus type 2 (Mad K87)			0/2		0/2		0/2	
Mouse cytomegalovirus					0/2		0/2	
Mouse hepatitis virus	0/2	MFI-serology	0/2	0/2	0/2	0/2	0/2	0/2
Mouse K-virus					0/2		0/2	
Mouse norovirus	2/2	MFI-serology	2/2	2/2	2/2	2/2	2/2	2/2
Mouse parvovirus	0/2	MFI-serology	0/2	0/2	0/2	0/2	0/2	0/2
Mouse polyoma virus					0/2		0/2	
Mouse rotavirus (EDIM)	0/2	MFI-serology	0/2	0/2	0/2	0/2	0/2	0/2
Mouse thymic virus					0/2		0/2	
Pneumonia virus of mice			0/2		0/2		0/2	
Reovirus type 3 (Reo 3)			0/2		0/2		0/2	
Sendai virus			0/2		0/2		0/2	
Theiler's murine encephalomyelitis virus	0/2	MFI-serology	0/2	0/2	0/2	0/2	0/2	0/2
BACTERIA, MYCOPLASMA AND FUNGI								
<i>Bordetella bronchiseptica</i>					0/2		0/2	
CAR Bacillus					0/2		0/2	
<i>Citrobacter rodentium</i>					0/2		0/2	
<i>Clostridium piliforme</i>					0/2		0/2	
<i>Corynebacterium kutscheri</i>					0/2		0/2	
<i>Helicobacter spp</i>	0/1*	PCR	0/1*	0/1*	0/2	0/1*	0/2	0/2*
<i>Klebsiella spp</i>					0/2		0/2	
<i>Mycoplasma pulmonis</i>			0/2		0/2		0/2	
<i>Pasteurella spp</i>	0/1*	PCR	0/1*	0/1*	0/2	0/1*	0/2	0/1*
<i>Proteus spp</i>					0/2		1/2	
<i>Pseudomonas aeruginosa</i>					0/2		0/2	
<i>Salmonella spp</i>					0/2		0/2	
<i>Staphylococcus aureus</i>					0/2		0/2	
<i>Streptobacillus moniliformis</i>					0/2		0/2	
Streptococci Beta-haemolytic (A, B, C, G)	0/1*	PCR	0/1*	0/1*	0/2	0/1*	0/2	0/1*
<i>Streptococcus pneumoniae</i>	0/1*	PCR	0/1*	0/1*	0/2	0/1*	0/2	0/1*
<i>Yersinia pseudotuberculosis</i>					0/2		0/2	
PARASITES								
Ectoparasites	0/1*	PCR	0/1*	0/1*	0/2	0/1*	0/2	0/1*
<i>Encephalitozoon cuniculi</i>					0/2		0/2	
Endoparasites	0/1*	PCR	0/1*	0/1*	0/2	0/1*	0/2	0/1*
PATHOLOGICAL LESIONS								
External	0/2**	Pathology	0/2**	0/2**	0/2	0/2**	0/2	0/2**
Necropsy	0/2**	Pathology	0/2**	0/2**	0/2	0/2**	0/2	0/2**

* 1 sample (tested by PCR): refers to a sample, containing samples from 10 animals (pooled) (2 sentinels plus 8 colony animals).

**BSRC Alexander Fleming

Signature of Veterinarian
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